

2-29-00

Theory of CC

Discuss calc. of yield, and m

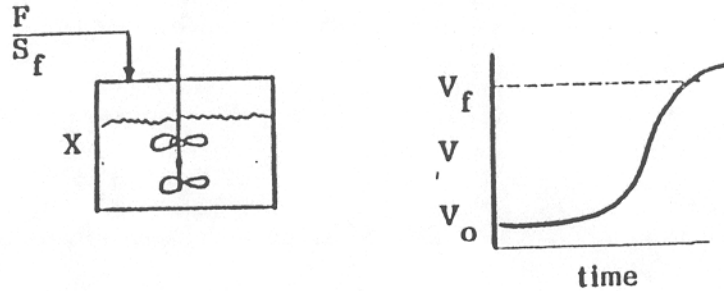
TOPIC 10:

THEORY AND APPLICATIONS OF CONTINUOUS CULTURE

BY:

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FED-BATCH FERMENTATION KINETICS



Growth in batch culture:

$$V \frac{d[X(t)]}{dt} = V \mu(t) [X(t)]$$

Fed-batch culture:

$$\frac{d[(X(t) V(t))]}{dt} = \mu(t) [X(t) V(t)]$$

$$\frac{d[V(t)]}{dt} = F_1(t) + \dots + F_n(t)$$

Substrate balance:

$$\frac{d[S(t) V(t)]}{dt} = F_s(t) S_f - \frac{\mu(t) [X(t) V(t)]}{Y(t)}$$

Operating objective: $\frac{dS(t)}{dt} = 0$ when $S(t)$ is near 0

Therefore $\frac{S(t) dV(t) + V(t) dS(t)}{dt} \approx 0$

And $F_s(t) = \frac{\mu(t) [X(t) V(t)]}{Y(t) S_f}$

COMPUTER CONTROL OF YEAST PRODUCTION

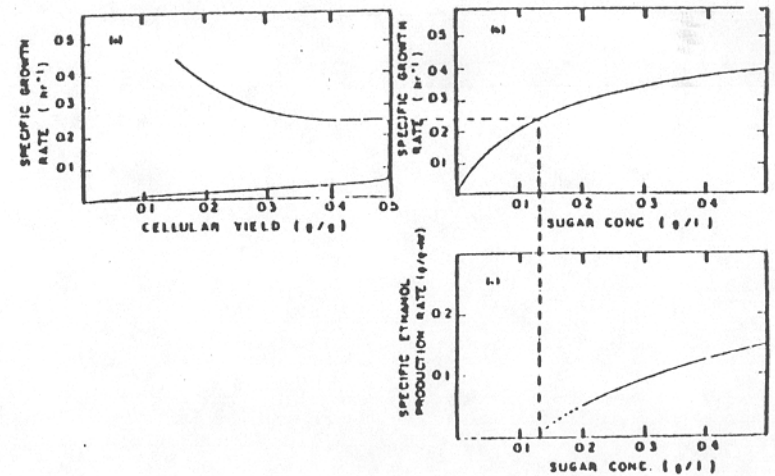
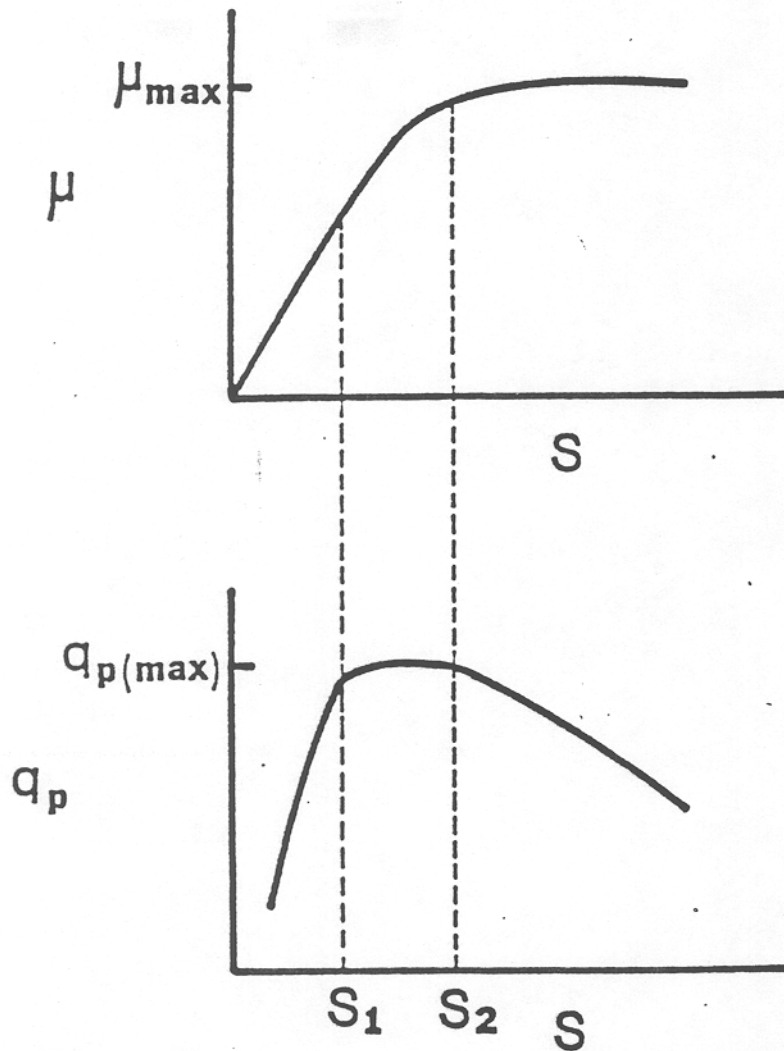


Fig. 3. Relationships between (a) specific growth rate and cell yield, (b) sugar concentration and specific growth rate, and (c) sugar concentration and ethanol production rate in a bakers' yeast fermentation.

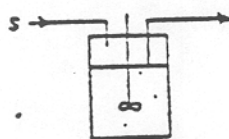
Microbial Growth and Product Formation



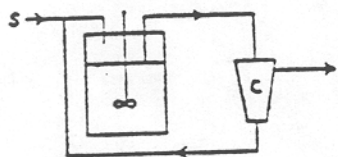
μ =specific growth rate
 S =substrate concentration
 q_p =specific productivity

Types of Continuous Culture Systems

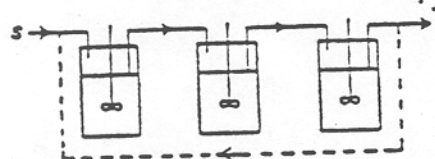
A. Homogeneous
1. Single stage
(i) Stirred fermentor



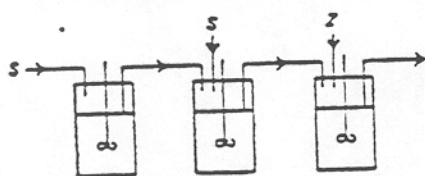
(ii) Stirred fermentor with feed-back



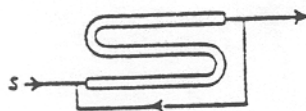
2. Multi-stage
(i) Simple chain



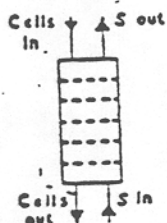
(ii) Multiple substrate addition



B. Heterogeneous
1. Single phase
(i) Pipe flow with feed-back



2. Multi-phase packed towers
(i) Liquid-liquid
(ii) Liquid-gas



C. Mixed
(i) Stirred tank feeding tubular reactor

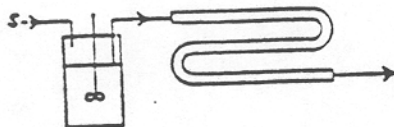
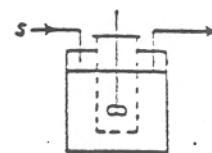
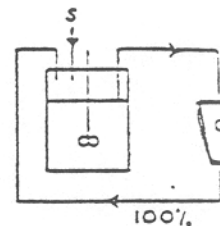


FIG. 1. Types of 'open' continuous culture systems
S indicates substrate addition, C indicates continuous centrifuge or settling tank

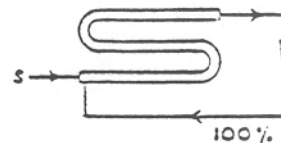
A. Homogeneous
(i) "Cellophane bag" cultures



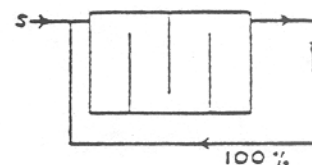
(ii) Stirred fermentor with 100% feed-back of cells



B. Heterogeneous
1. Single phase
(i) Pipe flow with 100% feed-back of cells



(ii) Partitioned tank with 100% feed-back of cells



2. Two-phase
(i) Pellicle growth



(ii) Packed towers

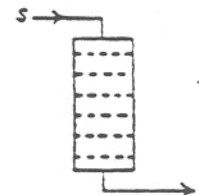


FIG. 2. Types of 'closed' continuous culture systems
S indicates substrate addition, C indicates continuous centrifuge or settling tank

Taken from Herbert (1961) Continuous Culture of Microorganisms
Society of Chemical Industries Monograph No. 12, pp. 21.

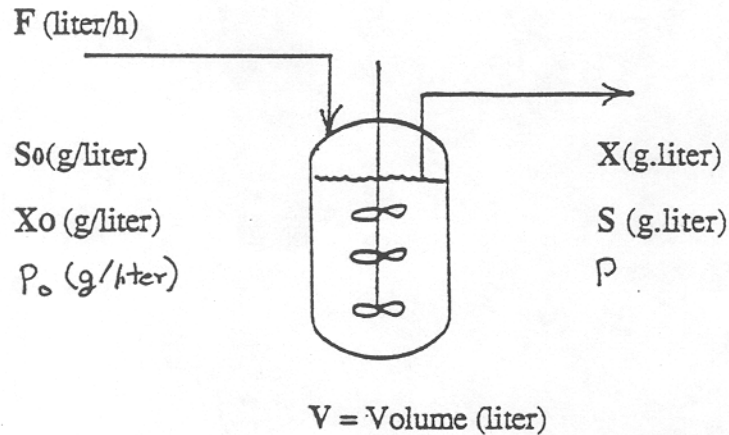
S

S

SUBSTRATE CONC.

3

Analytical Description of a Single-Stage Chemostat



Material balance on cell mass

Accumulation = Cells In - Cells Out + Growth - Lysis

$$\frac{VdX}{dt} = F X_0 - F X + \mu XV - \alpha XV$$

At steady state ($dX/dt = 0$), assume: Medium feed is sterile ($X_0 = 0$) and growth is much greater than lysis ($\mu \gg \alpha$)

$$\frac{VdX}{dt} = 0 = \mu XV - FX$$

$$\frac{F}{V} = D \text{ the Dilution Rate}$$

$$\mu = D$$

Specific growth rate
= Dilution rate

Material balance on limiting nutrient

Accumulation = Nutrient In - Nutrient Out - (Consumption)

$$\frac{VdS}{dt} = F S_0 - F S - \frac{\mu XV}{Y_{x/s}} - m XV - \frac{q_p XV}{Y_{p/s}}$$

Substrate Allocation Model

At steady state ($dS/dt = 0$), assume that $\mu/Y \gg m$ and $q_p = 0$

$$0 = F S_0 - F S - \frac{\mu X V}{Y}$$

Substituting $F/V = D$ and $D = \mu$

$$\Rightarrow X = Y (S_0 - S)$$

Assuming a growth model $\mu = f(S)$:

$$\mu = \mu_m \left(\frac{S}{K_s + S} \right)$$

then in continuous culture,

$$S = K_s \left(\frac{D}{D_c - D} \right)$$

and substituting

$$\Rightarrow X = Y \left(S_0 - K_s \frac{D}{D_c - D} \right)$$

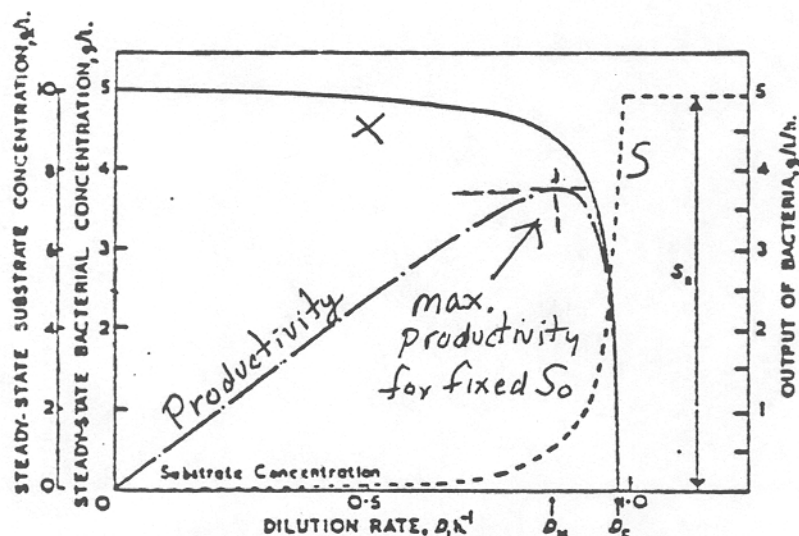
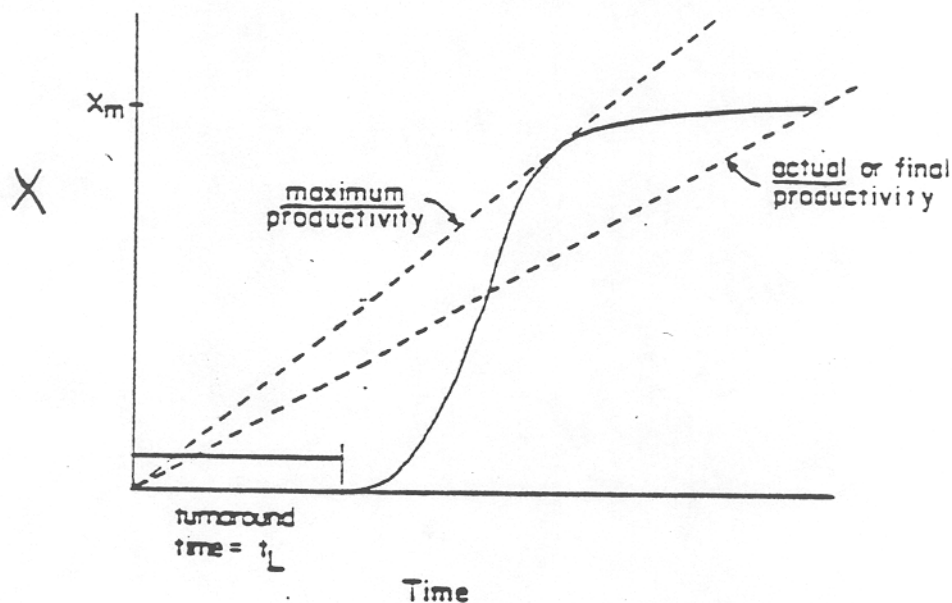


Fig. 2 Steady-state relationships in a single-stage continuous culture (theoretical). The steady-state values of substrate concentration, cell concentration and output at different dilution rates are calculated from equations (14) and (15) for an organism with the following growth constants: $\mu_m = 1.0 \text{ h}^{-1}$, $Y = 0.5$ and $K_s = 0.2 \text{ g/L}$; and a substrate concentration in the inflowing medium of $S_0 = 10 \text{ g/L}$.

— bacterial concentration
 --- output of bacteria
 - - - substrate concentration.

$$\begin{aligned} \mu_m &= 1 \text{ h}^{-1} \\ Y &= 0.5 \text{ g/g} \\ K_s &= 0.2 \text{ g/L} \\ S_0 &= 10 \text{ g/L} \end{aligned}$$



$$\text{Productivity} = \frac{\mu_m Y \left(\frac{x_m - x_0}{x_m} \right)}{\ln \frac{x_m}{x_0} + \mu_m t_L}$$

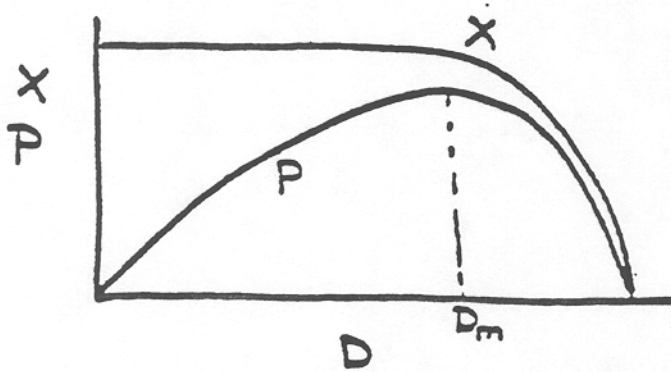
x_m = final cell mass
 x_0 = inoculum size
 t_L = turnaround time

(Herbert et al., 1956. J. Gen. Microbiol. 5:698)

Typical Turnaround Times:

	t_2	t_{ferm}
Yeast Fermentation	4- 8	18- 24
Antibiotic Fermentation	12-18	120-240

Productivity in Continuous Culture



$$\text{Productivity} = P = DX \quad (\text{g cell/liter-h})$$

$$\frac{\text{Continuous Productivity}}{\text{Batch Productivity}} = \ln \frac{x_m}{x_i} + t_1 \mu_{\text{max}}$$

x_m = max. cell density
 x_i = initial cell density
 t_1 = turnaround time
 μ_{max} = max. specific growth rate

ASSUMPTIONS IN THE DEVELOPMENT OF THE THEORY OF A SINGLE-STAGE CHEMOSTAT

1. Liquid volume, V , is a constant
2. Vessel is well-mixed
3. Single growth limiting nutrient, S_0
4. Steady state, $dX/dt = dS/dt = 0$
5. Medium feed is sterile, $X_0 = 0$
6. Growth is much faster than cell lysis, $\mu \gg \alpha$
7. No product formation, $q_p = 0$
8. Cell maintenance is low, $\mu/Y \gg m$
9. No mass or heat transfer limitations
10. Cell Yield, Y , is constant
11. Growth model, $\mu = F(s)$, e.g. the Monod Model

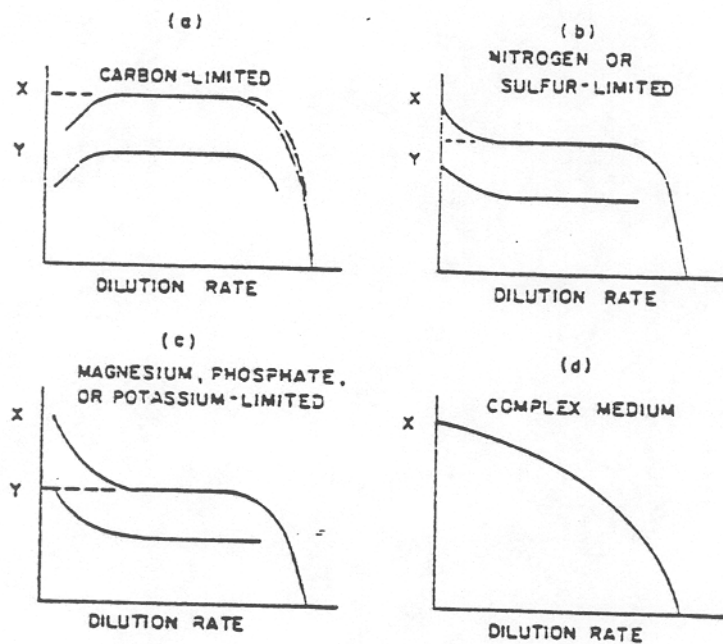
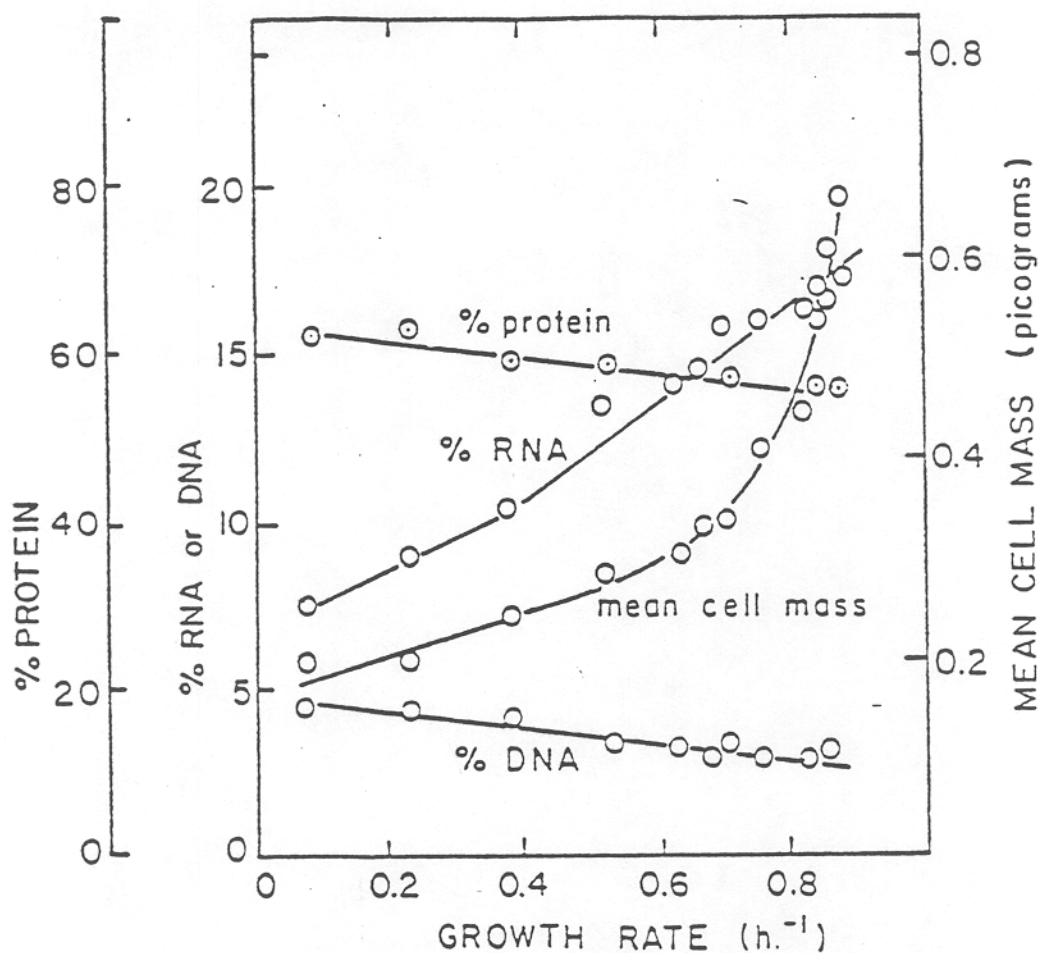


Figure 7.6 Examples of nonideal behavior of the single-stage, well-mixed chemostat; explanations are in the text. Dotted lines represent expected behavior.



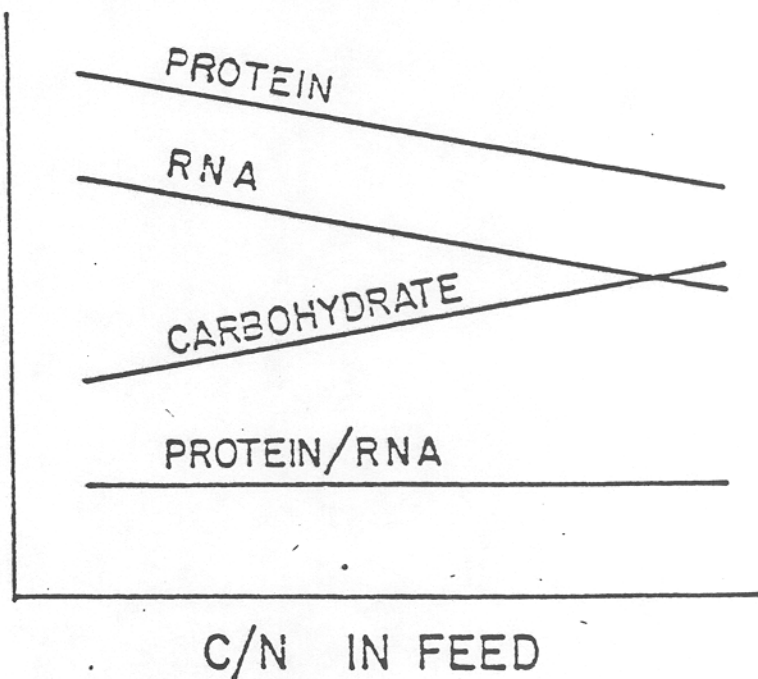
Cell composition of *Aerobacter aerogenes* under N-limitation at different growth (dilution) rates in continuous culture.

% PROTEIN

% RNA

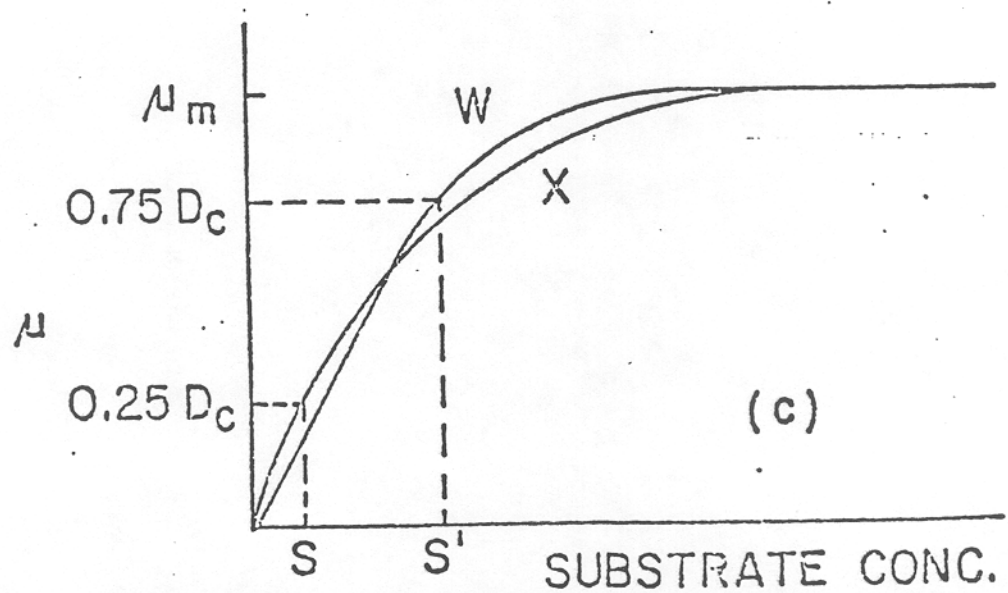
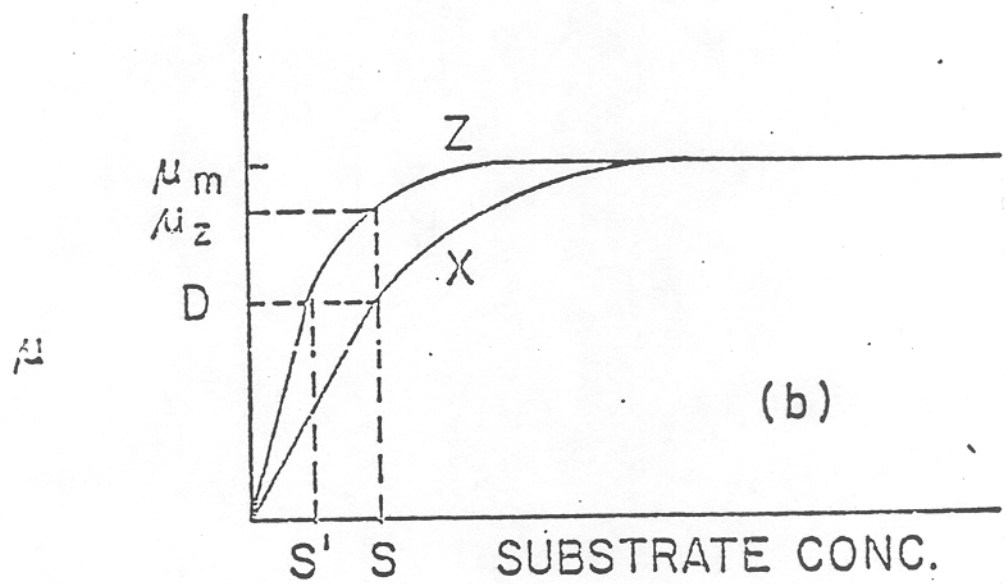
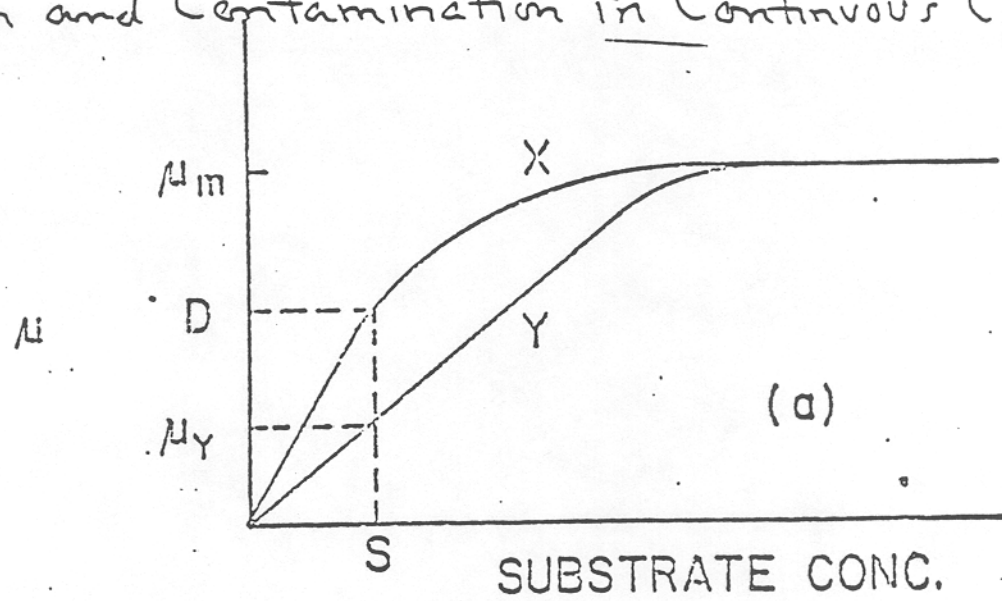
% TOTAL CARBOHYDRATE

PROTEIN/RNA

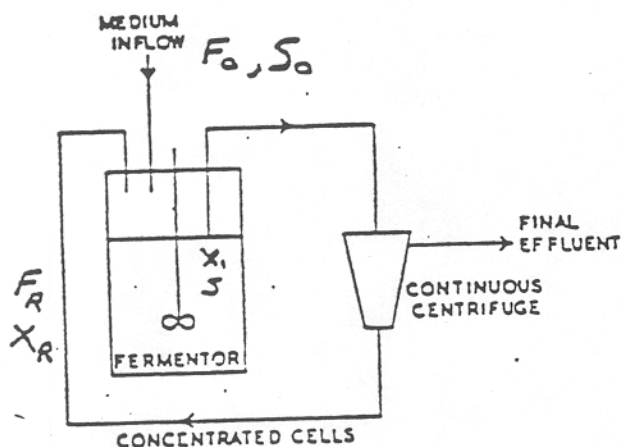


Variations of cell composition with dual nutrient limitation in continuous culture.

Selection and Contamination in Continuous Culture



CELL RECYCLE IN CONTINUOUS CULTURE



Single-Stage Stirred Fermentor with

Single-Stage Stirred Fermentor with
Feed-Back of Cells

Steady State Equations:

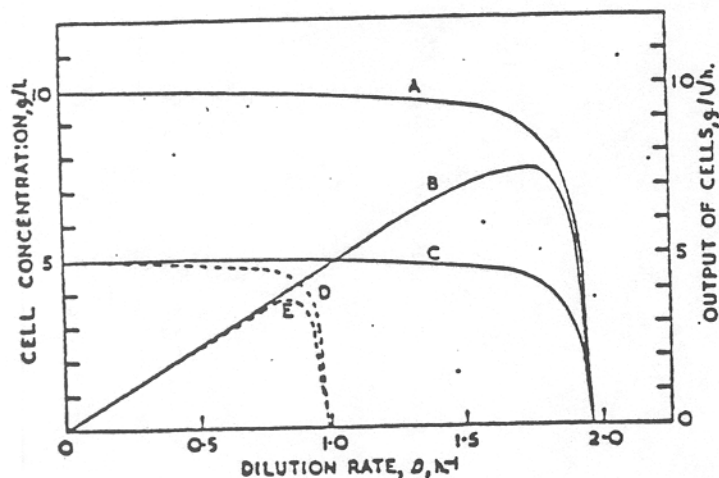
$$\mu = D [1 - \alpha (\beta - 1)]$$

$$\therefore \mu < D$$

$$X_1 = Y / (1 - \alpha (\beta - 1)) (S_0 - S)$$

$$\alpha = \frac{F_R}{F_0}$$

$$\beta = \frac{X_R}{X_1}$$

Steady-State Relationships in
A Single-Stage Continuous Cul-
ture with Feed-Back (Theoreti-
cal)

Curves are plotted for an organism with the following growth-constants: $\mu_m = 1.0 \text{ h}^{-1}$; $Y = 0.5$; $K_0 = 0.2 \text{ g/l}$; and a substrate concentration in the inflowing medium of $S_0 = 10 \text{ g/l}$. Continuous curves: with feed-back (volumetric feed-back ratio $\alpha = 0.5$; cell concentration factor $\beta = 2.0$). Dotted curves: without feed-back. Curve A, cell concentration in fermentor; Curve B, output of cells; Curve C, cell concentration in outflow; Curve D, cell concentration without feed-back; Curve E, output without feed-back.

CELL RECYCLE

REF: Gold, Mohagheghi, Cooney and Wang,
Biotech. Bioeng. 23:2105-2116 (1981)

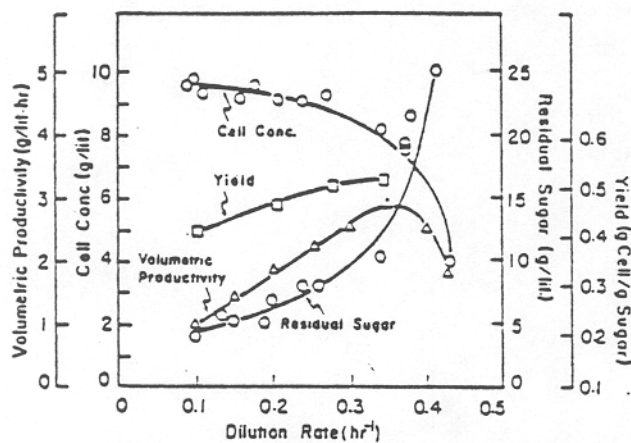


Fig. 3. Continuous culture of *C. utilis* on SSL at 30°C and pH 4.5.

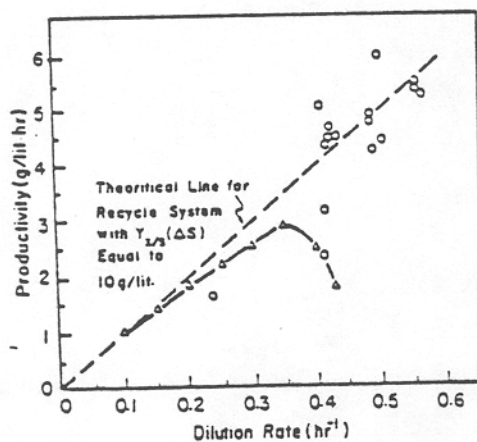


Fig. 5. Comparison of productivity for continuous culture (—) with and (---) without recycle.

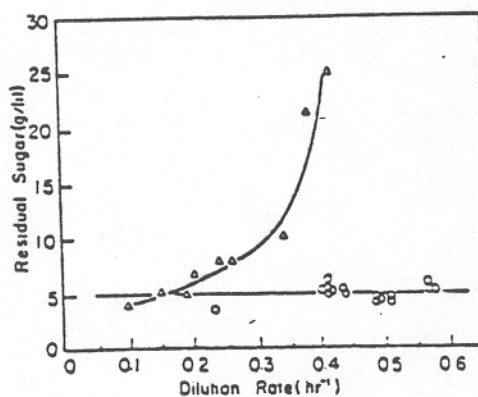
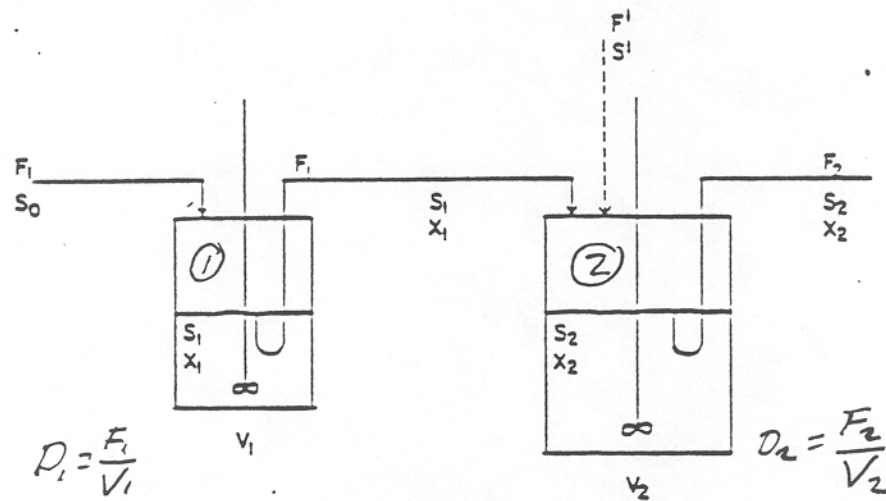


Fig. 6. Comparison of effluent sugar for continuous culture (Δ) with and (○) without recycle.

MULTISTAGE CONTINUOUS CULTURE



Steady State Solutions for Material Balances on
First and Second Stages

	Cell Mass Balance	Substrate Balance
First Stage	$\mu_1 = D_1$	$X_1 = Y (S_0 - S_1)$
Second Stage	$\mu_2 = D_2(1 - \frac{X_1}{X_2})$	$X_2 = \frac{D_2}{\mu_2} Y (S_1 - S_2)$
Second Stage with additional feed system	$\mu_2 = D_2 - \frac{F_1}{V_2} \frac{X_1}{X_2}$	$X_2 = \frac{Y}{\mu_2} \left(\frac{F_1 S_1}{V_2} + \frac{F S'}{V_2} - D_2 S_2 \right)$

PRODUCT ACCUMULATION IN CONTINUOUS CULTURE

PROBLEM:

Many investigators have proposed that penicillin be produced in continuous culture. To address this question, examine the effect of dilution rate on product concentration and yield.

DATA:

q_p	=	6 units penicillin G/mg-cell-h
X (steady state)	=	20 g/liter
m	=	0.022 g/g cell-h
$Y_{p/s}$	=	1.1 g/pen-G/g glucose
$Y_{x/s}$	=	0.5 g/g glucose

PRODUCTION OF PENICILLIN IN CONTINUOUS CULTURE

$$\text{Growth} \quad \frac{dX}{dt} = \mu X - DX$$

$$\text{Substrate} \quad \frac{dS}{dt} = D S_0 - D S - \frac{\mu X}{Y_{x/s}} - m X - \frac{q_p X}{Y_{p/s}}$$

$$\text{Product} \quad \frac{dP}{dt} = q_p X - D P$$

SUGAR CONSUMPTION (GM/L-HR)

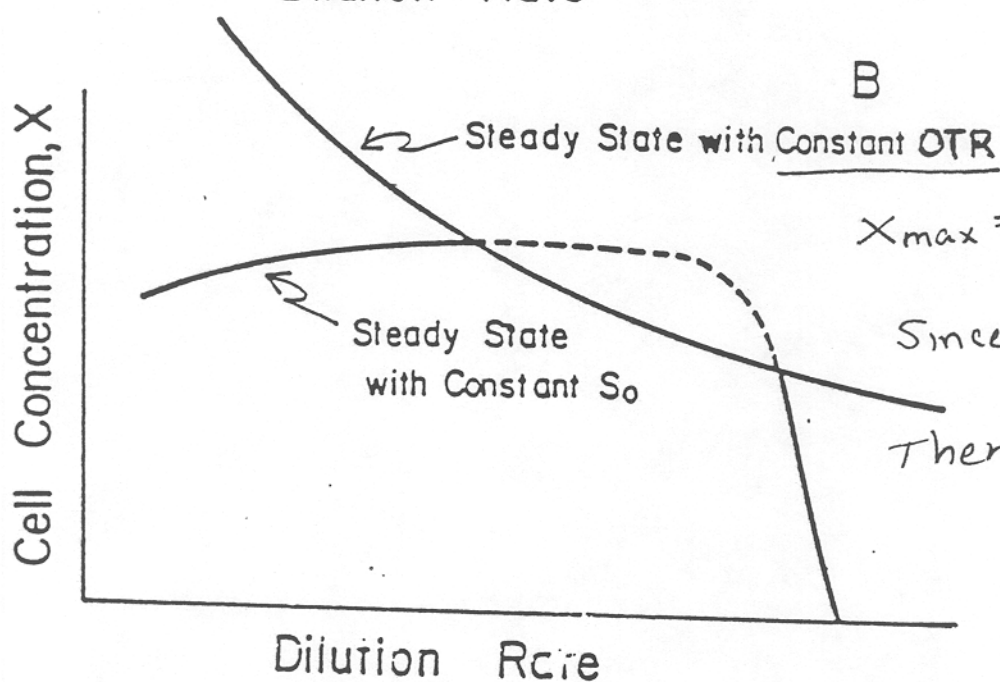
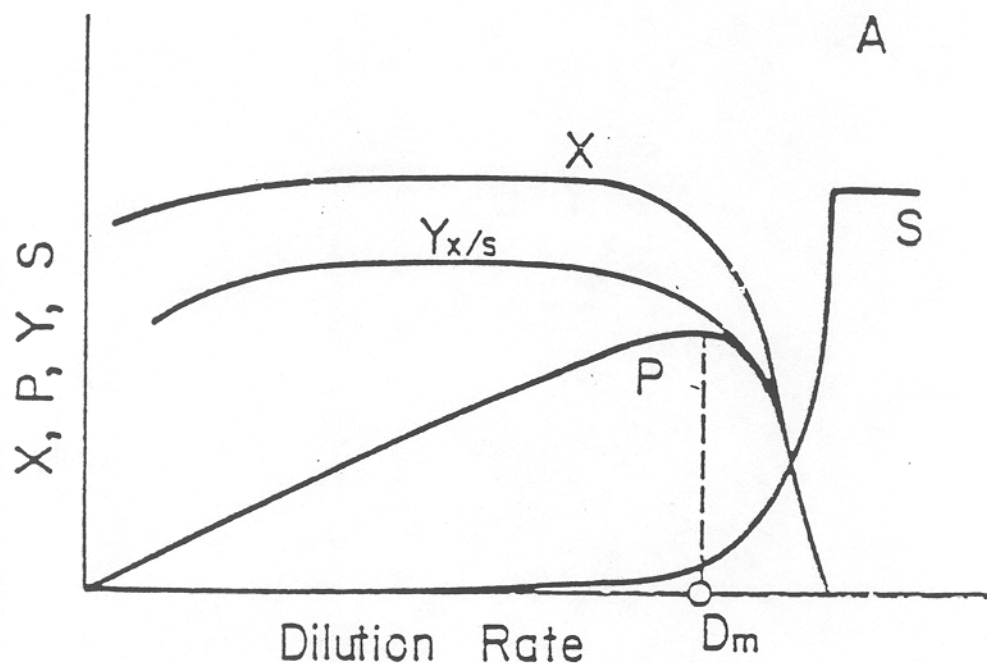
<u>Dilution Rate (1/h)</u>	<u>0.05</u>	<u>0.1</u>	<u>0.15</u>
Cell Mass	2	4	6
Maintenance	0.44	0.44	0.44
Penicillin	0.06	0.06	0.06
TOTAL	2.5	4.5	6.5

$$X = 20 \text{ g/liter}; \quad m = 0.022 \text{ g/g cell-h}$$

$$Y = 1.1 \text{ g pen/g glucose}; \quad Y_{x/s} = 0.5 \text{ g/g}$$

$$q_p = 6 \text{ units/mg cell-h}$$

Selection of Optimum Dilution Rate for Biomass Production



COMMERCIAL APPLICATION OF CONTINUOUS CULTURE

Food and Feed Processes

Single-Cell Protein:

Mycoprotein

Methanol

(ICI, Methylophilus methylotrophus)

Spent Sulfite Liquor

(Candida utilis)

Cheese whey

Molasses

n-Parafins

Wood hydrolyzate

Beverage:

Wine

Beer

Potable alcohol

Vinegar

Baker's Yeast

Yeast extract

Waste Water Treatment

Aerobic

Anaerobic

Single Stage

Multiple Stage - acidification/methanation

Chemicals

Gluconic Acid

Acetone/Butanol

Ethanol

Hyaluronic acid

Enzymes/Proteins

Glucose Isomerase

Diagnostic Enzymes

Insulin

Protein A

Immobilized Whole Cells

Aspartic Acid

Malic Acid

6-Aminopenicillanic Acid

Glucose Isomerization

Amino Acid Resolution

Ethanol

Waste Treatment systems

Animal Cell

tPA (Recombinant)

MAB (Hollow-Fiber)

TPA/MAB (microcarrier)