Dictyostelium amoebae are free living cells with a remarkable twist: under the stress of starvation, large numbers of amoebae are able to collect together to form a single multi-cellular organism (Fig. 1). The entire process begins when starving amoebae emit pulses of the chemoattractant cAMP, inducing the surrounding cells to move in their direction and to secrete cAMP themselves. This process generates outgoing spiral waves of cAMP which direct the entire population towards the original source (Fig. 2). We will try to understand the origin of and response to these cAMP waves.

1. **Chemical kinetics of cAMP signaling.**

Two species of cAMP receptors exist in the Dictyostelium cell membrane: an ‘activator’ $A$, and an ‘inhibitor’ $I$, both of which act on a third protein $R$. When bound to cAMP, a pair of $A$ molecules catalyzes the conversion of $R$ to an active form $R^*$, and a pair of $I$ molecules catalyzes the reverse reaction (Fig. 3).

The initial binding of cAMP (C) to its receptors is described by:

\[
A + C \xrightleftharpoons{k_A^+}{k_A^-} AC \quad \quad I + C \xrightleftharpoons{k_I^+}{k_I^-} IC
\]

Write down equations describing the time evolution of $[AC]$ and $[IC]$. Assume that $[AC] \ll [A_{tot}]$, and $[IC] \ll [I_{tot}]$; let $a$ be proportional to $[AC]$, $i$ to $[IC]$, and $c$ to $[C]$. By making a convenient choice of units, show that these equations can be written in the form

\[
\frac{da}{dt} = k_A^- (c - a) \quad \quad \frac{di}{dt} = k_I^- (c - i).
\]
b. The reactions involving activation and inactivation of $R$ reach a rapid equilibrium:

$$R \overset{k^+_R}{\underset{k^-_R}{\rightleftharpoons}} R^*$$

The $a^2$ and $i^2$ terms arise because it takes two molecules of $AC$ or $IC$ to catalyze these conversion reactions. Setting $\beta = k^-_R / k^+_R$, find an expression for the rapid equilibrium value of $r \equiv [R^*]$ in terms of $a$, $i$, and $[R_{tot}]$.

$$R_{tot} \equiv a^2 + i^2$$

2. Positive feedback and oscillations.

The molecule $R$ is an enzyme known as adenylate cyclase, which in its active form catalyzes the conversion of ATP into cAMP in the cytoplasm. The presence of extracellular cAMP thus stimulates the synthesis of intracellular cAMP, which in turn is secreted into the environment, creating a positive feedback loop (Fig. 3). Let $c_1 \equiv [\text{cAMP}_{in}]$; let the rate of cytoplasmic cAMP synthesis be $k_1 r$; and let the rate constant for its secretion be $k_0$. cAMP is continuously degraded by phosphodiesterase enzymes both inside and outside the cell, with rate constants $\gamma_1$ and $\gamma_0$, respectively. The entire network is described by the following equations:

$$\frac{dc_1}{dt} = k_1 r - (\gamma_1 + k_0)c_1 \quad \frac{dc}{dt} = k_0 c_1 - \gamma_0 c.$$  \hspace{1cm} (2)

a. Assuming that the concentrations $c_1$ and $a$ reach rapid equilibrium, reduce the four equations in (1) and (2) to two equations for the slow variables $c$ and $i$. Show explicitly the choice of units required to produce the simplified form shown in (3). What is the value of $k$?

$$\frac{dc}{dt} = \frac{c^2}{\beta \cdot i^2 + c^2} - c \quad \frac{di}{dt} = k(c - i).$$  \hspace{1cm} (3)

b. When *Dictyostelium* is grown in liquid medium, extracellular cAMP is well stirred, making its concentration uniform over space. However, under such conditions, the cAMP concentration is known to oscillate over time. Find the conditions on $k$ and $\beta$ so that the system is oscillatory.

c. On a graph of $i$ vs. $c$, plot the nullclines and simulate the time evolution of the system for an oscillatory case.

d. Suppose the system only required a single molecule of $AC$ or $IC$ in order to catalyze the $R$ conversion reactions. Comment on the behavior of the system in this case.
3. *Adding stochasticity to the system.*

In order to give the model a stochastic interpretation we will write a corresponding deterministic toy-model system of the form

\[
\frac{dn_c}{dt} \equiv \frac{\nu(h + n_c^2)}{h + h' + \beta \cdot n_i^2 + n_c^2} - n_c, \quad \frac{dn_i}{dt} \equiv \tilde{k}(\alpha n_c - n_i). \tag{4}
\]

Where \(n_c\) represents the number of molecules of \(c\) and \(n_i\) the number of molecules of \(i\). We have also included some leakage terms (associated with the terms involving \(h\) and \(h'\)) that transform \(R\) into \(R^*\) and vice versa in order to prevent the system from stalling in the case in which \(n_c = n_i = 0\). We will consider the choice of parameters \(\alpha = 1, \beta = 4; \tilde{k} = 0.1\) and \(h = h' = 1\).

4. *Diffusion and cAMP waves.*

When cells are grown on a plate, cAMP diffusion is slow, and the extracellular cAMP concentration is no longer uniform. Consider a plate on which there exists a uniformly distributed population of cells. Each cell senses cAMP in its environment, and secretes fresh cAMP in response. This new batch of cAMP is able to reach neighboring cells, stimulating them to synthesize more cAMP, and so on. The situation is similar to one in which a number of radio transmitter towers (cells) are used to detect, amplify, and re-broadcast a weak radio signal (cAMP). The cAMP concentration now varies over space as well as time. For simplicity, we will analyze a 1-dimensional case, with cells uniformly distributed along a line. We can assume that the cells have fixed positions over the timescales considered, because their chemotaxis is relatively slow. This system obeys the equations

\[
\frac{\partial c}{\partial t} = \frac{c^2}{\beta \cdot i^2 + c^2} - c + D \frac{\partial^2 c}{\partial x^2}, \quad \frac{\partial i}{\partial t} = k(c - i). \tag{10}
\]

In this problem you will have to develop a numerical code for simulating these equations. The system is assumed to extend from \(x = -1\) to \(x = +1\), with no flow at the boundaries. As for the initial conditions, consider a pulse of cAMP centered in the origin, that goes to zero towards the boundaries and that has a typical width \(\sigma\) (try to use a smooth but localized function); assume an uniform initial concentration of inhibitor \(i_0\).

10. a. Use the following parameters: \(\beta = 4; k = 0.5; D = 10^{-7}; \sigma = 0.1; i_0 = 0.1\). Run the simulation to see the emergence of cAMP waves emanating from the origin. Plot out a typical cAMP profile, indicating the direction of motion of the waves.
b. Run the simulation again, this time with a $k$ value which produces a non-oscillating system. Describe the typical cAMP profile once the transients have died out. Can cells find the initial source of cAMP based on this type of profile?

c. A simple concentration gradient would allow cells to find the cAMP source. Why do you think Dictyostelium uses waves of cAMP rather than a gradient in order to trigger cell aggregation?

5. **Receptor clustering and signal amplification.**

The cells must now sense and move towards the source of the cAMP waves. It is unknown precisely how the amoebae are able convert the small front-to-back difference in cAMP concentration into the large output signal necessary to drive cell motion. However, a similar amplification of small temporal changes in ligand concentration into large changes in receptor activity has been observed in bacterial chemotaxis networks. We will now discuss a receptor clustering model that has been proposed in order to explain this amplification. Consider a receptor molecule $R$ that is activated by the binding of some ligand $L$. Let $\alpha = 0, 1$ represent the activity of the receptor. Thus,

\[
\frac{k}{k-} \begin{array}{c} R + L \\ (\alpha=0) \end{array} \rightarrow RL \quad (\alpha=1)
\]

a. The probability that the system will be found in a state with energy $E$ is proportional to $e^{-E/kT}$.

Show that the energy of the receptor molecule may be written in the form

\[-E/kT = A \cdot \alpha.\]

By calculating the mean activity $\bar{\alpha}$ and equating this to the value known from chemical kinetics, find an expression for $A$ in terms of $L$, $k_+$ and $k_-$. 

b. Now consider a lattice of receptors $R_i$, each with activity $\alpha_i$. Suppose that an active receptor is able to activate nearby receptors even if they are not ligand-bound. The energy of $R_i$ can then be written as

\[-E_i/kT = A \cdot \alpha_i + B \cdot \sum_{j=1}^{n} (\alpha_i - \frac{1}{2})(\alpha_j - \frac{1}{2})\]

where the sum over $j$ is a sum over the $n$ nearest neighbors of $R_i$. How does the state of the neighboring receptors influence the energy of $R_i$?

c. Assume that the activity of each neighboring receptor may be approximated by its mean value. That is, $\sum_{j=1}^{n} \alpha_j \equiv n \bar{\alpha}$. By substituting this expression into the above equation, find an expression for the mean activity $\bar{\alpha}_i$ of receptor $R_i$.

d. There is nothing special about the particular receptor $R_i$: our calculations could equally have been applied to any other receptor in the system. Therefore, the mean activity $\bar{\alpha}_i$ must be equal to the mean activity $\bar{\alpha}$ of the neighboring receptors. Apply this consistency condition to find an equation for $\bar{\alpha}$, and show how this equation may be solved graphically. (Hint: it is easier to work with the variable $s = \bar{\alpha} - 1/2$.)
(10) **e.** Explore the possible system responses as a function of the parameters \(A\) and \(B\). For low values of \(B\), the equation has a single solution for all \(A\) values. For high values of \(B\), the system goes from having one, to three, then back to one solution as \(A\) is swept from \(-\infty\) to \(+\infty\). Find the critical value \(B_c\) which separates these two behaviors. Explain why the system response changes as this critical value is crossed. Which regime is more relevant for understanding signaling?

(4) **f.** Assuming \(B < B_c\), calculate the logarithmic amplification

\[
G = \frac{\partial \ln(\alpha)}{\partial \ln(L)} \bigg|_{L_0} = \frac{L \cdot \partial \alpha}{\alpha \cdot \partial L} \bigg|_{L_0},
\]

where \(L_0 = \frac{k}{k_+}\) is the dissociation constant for ligand-receptor binding. Plot \(G\) as a function of \(B\), and verify that it approaches the correct limit as \(B\) approaches zero. We see that the amplification can be made arbitrarily large by appropriately tuning \(B\). What is the possible disadvantage of doing this?

(10) **g.** Based on published experimental data, estimate the logarithmic amplification at each stage of the *Escherichia coli* chemotaxis network. Is there any evidence that receptor clustering contributes to this amplification?