

## REVIEW SUMMARY

## NEURODEVELOPMENT

# Genes, circuits, and precision therapies for autism and related neurodevelopmental disorders

Mustafa Sahin\* and Mriganka Sur\*

**BACKGROUND:** Neurodevelopmental disorders are caused by abnormalities in the developing brain. Such abnormalities can occur as a result of germline or somatic mutations or because of epigenetic or environmental factors. These disorders affect a large number of children in the developed world, as well as the developing world. The societal cost of neurodevelopmental disorders is immense, making the pursuit of treatments for individuals with neurodevelopmental disorders a top unmet medical need.

**ADVANCES:** Research in the genetics of neurodevelopmental disorders such as autism suggests that several hundred genes are likely involved as risk factors for these disorders. This heterogeneity presents both a challenge and an opportunity for researchers. Although the exact identity of many of the genes remains to be discovered, functional analysis of genes underlying several single-gene disorders has yielded considerable progress. Most genes identified to date appear to encode proteins that serve certain

conserved pathways: protein synthesis, transcriptional or epigenetic regulation, and synaptic signaling. Genetic syndromes such as fragile X syndrome, Rett syndrome, and tuberous sclerosis complex provide insights into the molecular pathways commonly affected in autism spectrum disorder (ASD). Understanding the basic biology of these diseases has led to mechanism-based treatment designs.

These genetic disorders, once thought to be irreversible, are now the subject of trailblazing new clinical trials for neurodevelopmental disorders. On the basis of research in genetic mouse models, it is hypothesized that different genetic disorders will respond to different therapies, such as mammalian target of rapamycin inhibitors (tuberous sclerosis and PTEN hamartoma tumor syndrome), metabotropic glutamate receptor 5 antagonists (fragile X and 16p11.2 deletion), and insulin-like growth factor 1 (Rett and Phelan-McDermid syndromes). It is not yet clear whether such trials will result in approval of the drugs for these specific conditions. Subsets of non-syndromic autism patients may also benefit from

one of these therapies, but further investigation will be required to provide the tools and methods to stratify the individuals with non-syndromic autism into treatment groups.

A remaining hurdle is the lack of precise understanding about the brain regions and neuronal circuits underlying autism. Studies in mouse models of autism suggest abnormalities in specific brain regions, as well as in certain cell types. Excitatory and inhibitory neurons in the neocortex, as well as subcortical structures such as basal ganglia and cerebellum, have been implicated. Astrocytes and microglia also play roles in ASD. Further studies will be required to provide definitive evidence that similar brain regions, cell types, and circuits are relevant to autism symptoms in the human brain.

**OUTLOOK:** The next generation of research in neurodevelopmental disorders must address the neural circuitry underlying behavioral symptoms and comorbidities, the cell types in these circuits, and common signaling pathways that link diverse genes. Early attempts at treating neurodevelopmental disorders have yielded mixed

**ON OUR WEB SITE**

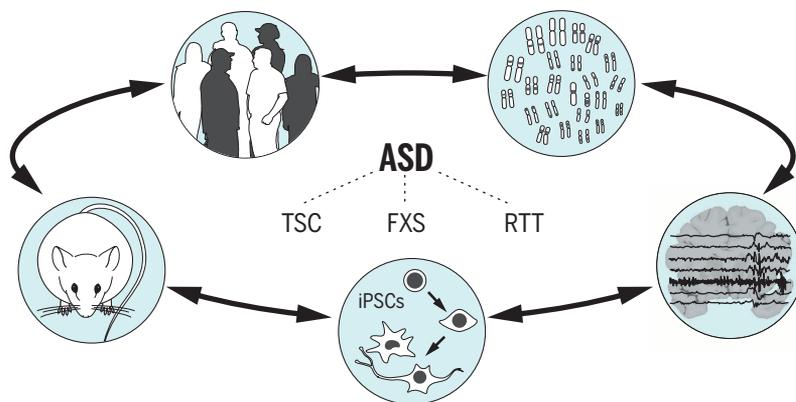
Read the full article at <http://dx.doi.org/10.1126/science.aab3897>

results, underscoring the necessity of choosing the right cohort of patients to treat, developing more sensitive and dynamic outcome measures, using cogent biomarkers, and utilizing technologies such as stem cell-derived neurons to predict response to treatment.

Biomarkers can be helpful in predicting subjects most likely to respond, confirm target engagement, and detect early signals of efficacy. Given that ASD represents circuit dysfunction, biomarkers that allow us to analyze autism-related circuit function are likely to be most relevant. Especially, translatable biomarkers that can be used in both mouse models and human subjects, such as electroencephalography, magnetic resonance imaging, visual or auditory evoked potentials, and eye-blink conditioning, can be particularly powerful.

One potential new tool to identify those who are likely to respond is induced pluripotent stem cell (iPSC)-derived neurons. This technology allows the possibility of testing the effects of a compound on a patient's neurons before it is given to the patient. Modeling the effects of mutations in iPSC-derived neurons can be informative about the molecular and cellular defects underlying autism.

Only when we can leverage the heterogeneity of neurodevelopmental disorders into precision medicine will the mechanism-based therapeutics for these disorders start to unlock success. ■



**Translational research and clinical trials in ASD.** Translational studies in ASD have gained momentum from genetically defined causes such as fragile X syndrome (FXS), Rett syndrome (RTT), and tuberous sclerosis complex (TSC). The patients with these disorders are phenotyped in detail by means of advanced imaging and electrophysiology studies, with the aim of identifying potential biomarkers. There are cell-based models (both rodent and human) as well as mouse models of these syndromes, enabling preclinical trials. Together, these efforts have led to clinical trials in some of these disorders. It is important to remember that the discovery cycle will likely take more than one round to achieve safe and effective therapies for these disorders.

The list of author affiliations is available in the full article online.  
\*Corresponding author. E-mail: [mustafa.sahin@childrens.harvard.edu](mailto:mustafa.sahin@childrens.harvard.edu); [msur@mit.edu](mailto:msur@mit.edu)  
Cite this article as M. Sahin and M. Sur, *Science* 350, aab3897 (2015). DOI: 10.1126/science.aab3897

## REVIEW

## NEURODEVELOPMENT

# Genes, circuits, and precision therapies for autism and related neurodevelopmental disorders

Mustafa Sahin<sup>1\*</sup> and Mriganka Sur<sup>2,\*</sup>

Research in the genetics of neurodevelopmental disorders such as autism suggests that several hundred genes are likely risk factors for these disorders. This heterogeneity presents a challenge and an opportunity at the same time. Although the exact identity of many of the genes remains to be discovered, genes identified to date encode proteins that play roles in certain conserved pathways: protein synthesis, transcriptional and epigenetic regulation, and synaptic signaling. The next generation of research in neurodevelopmental disorders must address the neural circuitry underlying the behavioral symptoms and comorbidities, the cell types playing critical roles in these circuits, and common intercellular signaling pathways that link diverse genes. Results from clinical trials have been mixed so far. Only when we can leverage the heterogeneity of neurodevelopmental disorders into precision medicine will the mechanism-based therapeutics for these disorders start to unlock success.

Neurodevelopmental disorders include a wide range of conditions such as epilepsy, intellectual disability, and autism spectrum disorder (ASD). Patients with ASD exhibit early-childhood onset of symptoms, first described over 60 years ago (1), that persist throughout life and produce notable impairments in social, communicative, cognitive, and behavioral functioning (2). According to the U.S. Centers for Disease Control, ASD affects 1 in 68 children and 1 in 42 boys. ASD is a major public health problem that leads to considerable disability and disrupts families, resulting in a total annual societal cost of ~\$35 billion in the United States alone (3).

ASD diagnosis comprises a constellation of behavioral symptoms, as defined by a group of experts (DSM-5), and requires persistent deficits in social communication and interaction across multiple contexts, as well as restricted, repetitive patterns of behavior, interests, and activities. A key characteristic in ASD is its heterogeneity. Patients with ASD present with wide variation and levels of impairment with different comorbidities, and the expression of these symptoms can change over time. Heterogeneity has been a huge obstacle in ASD research, but in recent years, researchers have started to take advantage of the heterogeneity of ASD. Rather than focusing on “pure autism” (autism not confounded by

intellectual disability) (4, 5), research has now opened up to examining genetic disorders with high penetrance of ASD, such as fragile X syndrome (FXS), Rett syndrome (RIT), and tuberous sclerosis complex (TSC), which have now come to the forefront of translational efforts to find treatments for subsets of mechanism-based classification of ASD (6). Complementary to this effort is the National Institute of Mental Health (NIMH) initiative to define psychiatric disorders according to mechanistic descriptions of symptom clusters rather than symptom inventories, also known as research domain criteria (7, 8). In ASD, the etiology seems to vary according to the individual's genome and interaction with his or her environment. Genetic heterogeneity and overlap with other neuropsychiatric disorders make it difficult to find a unique risk factor for ASD. Improved understanding and classification of ASD-based domains and levels of analysis could improve precision and treatment efficacy.

Here, we review research on neurodevelopmental disorders that spans genes, molecules, cells, and circuits, as well as the whole individual and environment. We discuss current efforts and obstacles in clinical trials and offer recommendations for the future that lead toward precision medicine.

## Genes

The genetic component of ASD susceptibility is evidenced by twin studies that demonstrated higher concordance of ASD among monozygotic than dizygotic twins and has benefited from modern genome scanning initiatives to yield many new genes worthy of further study. Genome analysis revealed the association of copy-number variants (such as 15q11-13, 16p11.2, and 22q11.2) and

single-nucleotide variants with ASD. Some of these variants are de novo (not found in either parent) and thus easier to deem as causal. Variants that are not de novo or sequencing variants that are not obviously deleterious are harder to evaluate. Several studies have used whole-exome sequencing to reveal a number of ASD susceptibility genes, such as *CHD8*, *GRIN2B*, and *SCN2A*. These studies estimate that 400 to 1000 genes are involved in ASD susceptibility (9). The vast majority of ASD susceptibility genes have not yet been identified and will require much larger cohorts for adequate statistical power, as was necessary for schizophrenia (10). Germline mutations are not the only contributor to brain disorders; somatic mutations that affect a subset of brain neurons can cause epilepsy, brain malformations, and quite possibly ASD (11). Somatic mosaicism affecting the brain will confound the genetic analysis of cohorts, which are almost always based on bulk DNA derived from the blood and intended to represent the inherited genome.

Along with larger cohort sizes, identifying many of the remaining hundreds of ASD susceptibility genes will require thoughtful and innovative study designs. One approach is to study families with consanguinity to reduce inherited variation and help identify rare recessive variants (12, 13). Another approach is to study groups that are relatively protected from ASD. Because ASD is much more common among males than females, focusing on families with a history of severe autism among women appears to enrich for highly penetrant rare variants (14).

The estimated heritability of ASD is 0.7 to 0.8, which, while relatively high, leaves room for non-inherited factors, including de novo mutations and epigenetic and environmental factors, leading to a complex risk architecture. Environmental influences such as perinatal injury and maternal infection could play an important role in the context of a susceptible genetic background and contribute to the development of ASD. For instance, premature infants with isolated cerebellar hemorrhage have a 30-fold higher incidence of ASDs compared to the general population (15, 16). Other epidemiological studies have implicated activation of the maternal immune system during gestation as a contributor to the development of various neuropsychiatric disorders (17–19) and more specifically in the development of autism (19–21). Maternal immune activation leads to region-specific changes in brain cytokines (22) and neuropathological changes that can be detected even in nonhuman primates (23). Interestingly, maternal immune activation is implicated in the exacerbation of syndromic forms of ASD. For example, maternal immune activation has been shown to intensify social behavior deficits observed in *Tsc2+/-* mutant mice (24). Finally, the relationship between the gut microbiome and neurodevelopmental symptoms has attracted attention (25). Autism and accompanying gastrointestinal symptoms are associated with distinct gut microbial compositions (26). Furthermore, probiotic treatment can improve both the metabolic abnormalities and the behavioral deficits in

<sup>1</sup>F. M. Kirby Center for Neurobiology, Translational Neuroscience Center, Department of Neurology, Boston Children's Hospital, Boston, MA 02115, USA. <sup>2</sup>Simons Center for the Social Brain, Picower Institute for Learning and Memory, Department of Brain and Cognitive Sciences, Massachusetts Institute of Technology, Cambridge, MA 02139, USA.

\*Corresponding author. E-mail: mustafa.sahin@childrens.harvard.edu; msur@mit.edu

a maternal immune activation mouse model, supporting a connection between gut microbiome and autism (27). Further studies are needed to test how robust these initial observations are and what cellular mechanisms mediate them.

**Molecular and cellular pathways**

Every identified ASD susceptibility gene sheds new light on the cellular mechanisms underlying ASD. Many of the genes implicated in ASD converge onto a few major signaling pathways: transcriptional control and chromatin remodeling, protein synthesis and cellular metabolism, and synapse development and function (6, 28–30). Although many of these cellular processes are shared between neurons and nonneuronal cells, they appear to play roles particularly relevant to ASD in the brain (Fig. 1).

**Transcriptional control and chromatin remodeling**

Several ASD genes influence transcription (31, 32), including those that are highly penetrant such as

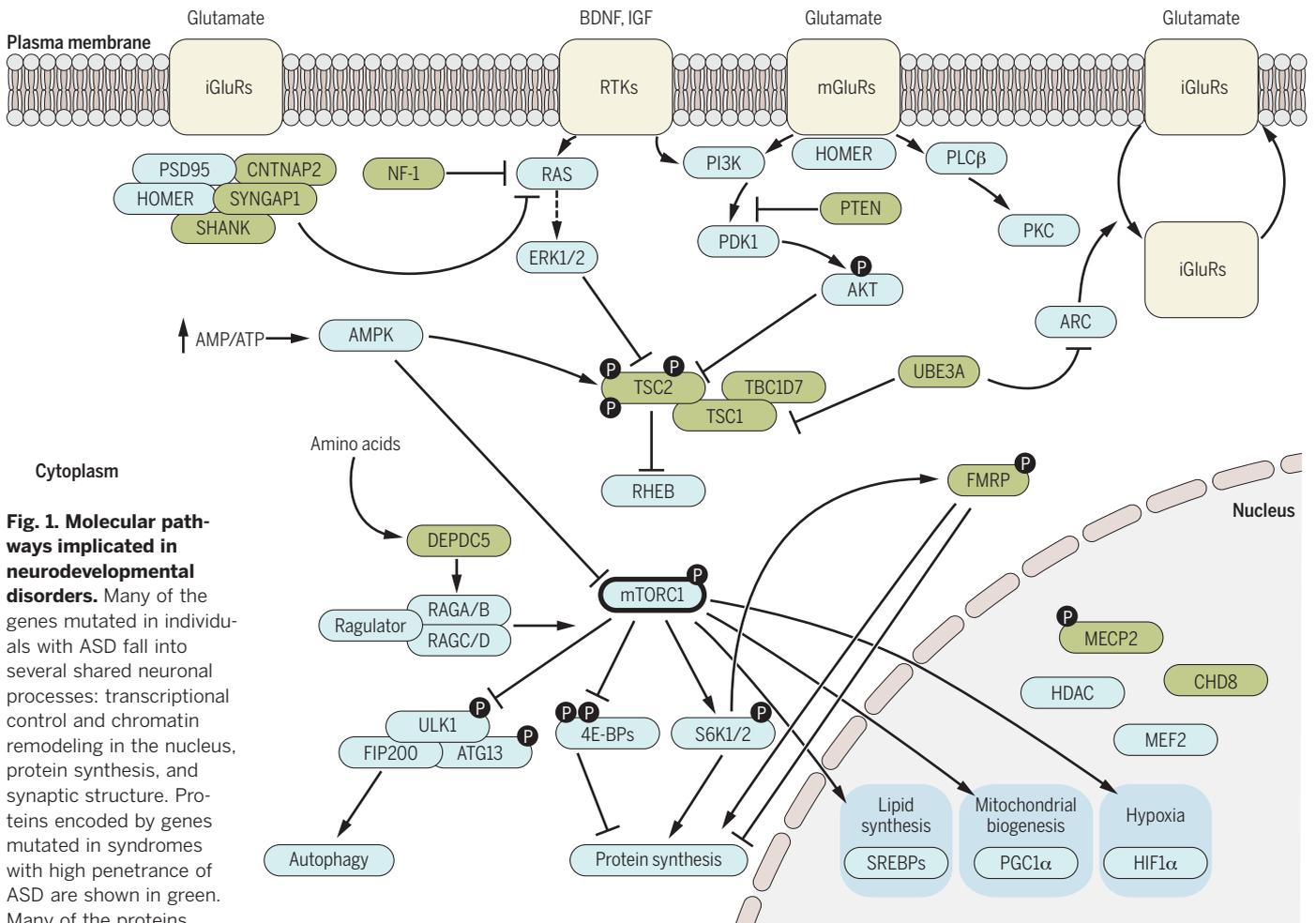
Rett syndrome. MeCP2 (methyl CpG binding protein 2), which underlies Rett syndrome, is a molecular multitasker that regulates gene expression by interacting with chromatin remodeling, transcription, and splicing (33). Initial findings suggested that MeCP2 binds to methylated CpG sites in the promoters of genes and associates with chromatin silencing complexes to repress gene expression (34–36). However, subsequent studies have demonstrated that MeCP2 also interacts with chromatin and transcriptional activators to activate gene expression (37, 38). MeCP2 also mediates microRNA-mediated posttranscriptional control of gene expression (39, 40), as does FMRP (fragile X mental retardation protein) (41).

**Signaling pathways and protein synthesis**

One cellular process that has been implicated in multiple studies is that of mRNA synthesis and protein translation (28). Two key pathways of protein synthesis that contribute to synaptic function are the PI3K/mTOR (phosphatidylinositol 3-kinase/mammalian target of rapamycin) pathway

and the Ras-MAPK (mitogen-activated protein kinase) pathway. These pathways have been linked to neurodevelopmental disorders and to synaptic dysfunction. Owing to the availability of specific and Food and Drug Administration (FDA)-approved inhibitors, the mTOR pathway has been well characterized. TSC and PTEN (phosphatase and tensin homolog) hamartoma tumor syndrome (PHTS) are two paradigmatic “mTORopathies” such that loss of TSC1, TSC2, or PTEN function leads to activation of mTOR kinase activity and high incidence of intellectual disability, seizures, and ASD (42). Other mutations in this pathway that present with ASD include the *neurofibromin 1 (NF1)* gene that results in neurofibromatosis type I. *NF1* encodes a guanosine triphosphatase-activating protein that suppresses the activity of the proto-oncogene *Ras* and also alters mTOR activity.

Dysregulation of protein synthesis is a prominent feature of several other neurodevelopmental disorders, such as FXS (43). FMRP is an mRNA binding protein that regulates the translation of mRNAs and is silenced in FXS, resulting in



**Fig. 1. Molecular pathways implicated in neurodevelopmental disorders.** Many of the genes mutated in individuals with ASD fall into several shared neuronal processes: transcriptional control and chromatin remodeling in the nucleus, protein synthesis, and synaptic structure. Proteins encoded by genes mutated in syndromes with high penetrance of ASD are shown in green. Many of the proteins (such as MECP2 and

FMRP) have multiple functions and interactions in the cell but are represented with the dominant functional role for the sake of clarity. Abbreviations not found in text include the following: RTKs, receptor tyrosine kinases; iGluRs, metabotropic glutamate receptors; PGC1-α, peroxisome proliferator-activated receptor gamma coactivator 1-alpha); SREBP, sterol regulatory element-binding proteins; HIF1α, hypoxia-inducible factor 1 alpha; ULK1, unc-51-like kinase 1; ARC, activity-regulated cytoskeleton-associated protein; UBE3A, ubiquitin protein ligase E3A.

aberrant protein synthesis from key transcripts implicated in synaptic plasticity (44). Likewise, while MeCP2 influences expression of several hundred genes (37), levels of BDNF (brain-derived neurotrophic factor) and IGF1 (insulin-like growth factor 1) are reduced in *Mecp2* mutant mice, along with other molecules that cause both the PI3K/mTOR and ERK/MAPK pathways to be down-regulated (37, 45–49). Treatment with recombinant human IGF1 up-regulates these pathways in mice and induced pluripotent stem cell (iPSC)-derived human neurons (50, 51) and ameliorates symptoms in mice (48). Preliminary results in human trials also appear promising (52). It is important to remember that PI3K/mTOR and ERK/MAPK pathways regulate a large number of cellular processes, including transcription, autophagy, metabolism, and organelle biogenesis and maintenance. The role of each of these cellular processes in the pathogenesis and therapeutics of ASD remains to be determined.

Disruptions of signaling pathways can change scaffolding of proteins at synapses. Such changes may cause neurodevelopmental disorders. PSD95 (postsynaptic density protein 95) anchors *N*-methyl-D-aspartate (NMDA) and AMPA receptors at glutamatergic synapses. PSD95 expression is influenced by PI3K signaling; its levels, as well as excitatory synaptic transmission, are reduced in *Mecp2* mutant mice (48) and rescued by IGF1 application. Similarly, *SHANK3*, which lies in the 22q13.3 deletion region associated with Phelan-McDermid syndrome (PMS), encodes a synaptic protein that regulates other protein partners, such as PSD95; up-regulation of the PI3K pathway by IGF1 rescues synaptic deficits in iPSC-derived human patient neurons (53) and *Shank3* mutant mice (54), at least partly by up-regulating PSD95.

Molecular convergence of pathways implicated by human genetics of ASD is apparent in studies of the *Fmr1* knockout mouse. First, among the

mRNA binding partners of FMRP are postsynaptic proteins such as SHANK3 and signaling proteins such as TSC2 and PTEN. Second, a number of studies in *Fmr1* knockout mice indicate that interfering with protein synthesis in different ways can normalize the phenotype of the knockout mice. Knockout of S6 kinase (55), Cebp (56), and PI3K (57, 58) is sufficient to ameliorate aspects of the *Fmr1* knockout pathogenesis, raising the possibility of having multiple potential targets for intervening with loss of FMRP. Dysregulation of metabotropic glutamate receptor (mGluR) and aberrant mGluR-dependent long-term depression (LTD) have been reported first in FXS mouse models (59, 60) and in several other ASD animal models, including *Nlgn3* (*neuroligin 3*) knockout and 16p11.2 knockout (61, 62). Although we do not yet know if murine hippocampal LTD models the human ASD brain function, these findings raise the possibility that convergent cellular and molecular pathway targets exist in subsets of ASDs.

### Brain regions and neural circuits

Molecular pathways in brain cells affect the function of neurons and synapses, and hence neuronal connectivity and circuits, to modify brain function. However, we lack insight about the brain regions and neuronal circuits underlying ASD. We do not yet know whether one cell type or circuit is crucial for the behavioral deficits observed in ASD patients. It is likely that different gene mutations perturb the neural circuitry underlying social interactions and repetitive behaviors at different nodes, resulting in a complicated matrix of genes, brain regions, and behavioral correlates (Fig. 2).

Histopathological and imaging-based evidence that implicates specific brain regions and circuits underlying ASD is limited. The pathological studies are hampered by small sample size, and thus there is an urgent need for systematic and wide-

spread collection of pathological specimens from those affected with a wide range of ASD. Imaging studies have been performed mostly on those with high-functioning ASD because patients must be able to tolerate and comply with magnetic resonance imaging (MRI) protocols. Thus, although the functional MRI studies performed to date implicate certain areas of the brain in the “high functioning” ASD group, it is not clear whether the same circuits are involved in those who have more severe cognitive deficits. It is also possible that differences between ASD and control groups identified in such studies do not represent the aberrant circuits that are causally related to the behavioral abnormalities, but instead represent the activation of other brain regions that compensate for the neural circuitry abnormalities. Clinical protocols that enable MRI studies in ASD patients with intellectual disability and those who are younger will enhance our understanding of ASD and its associated neural circuitry. Such studies performed in individuals with genetically identified subsets of ASD may also shed light on genotype-phenotype correlations (63, 64).

In addition to functional MRI, complementary techniques to investigate neuronal connectivity such as structural MRI, diffusion tensor imaging tractography, near-infrared spectroscopy, magnetoencephalography, and EEG can contribute to our understanding of brain connectivity at different time scales and with different spatial resolutions. It is likely that we will need to corroborate the findings from one modality with that from others to determine the most robust connectivity abnormalities in ASD. Some of these techniques may be more amenable for individuals at different ages and at different functional levels.

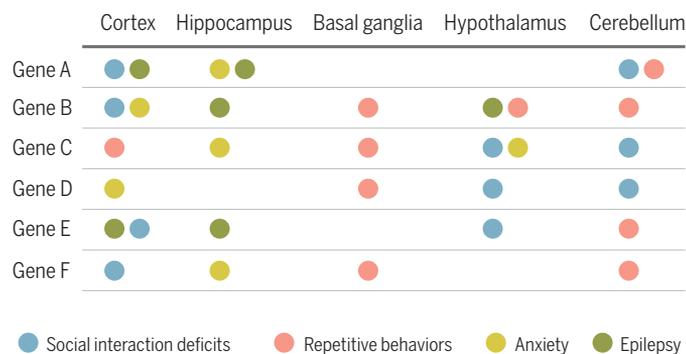
Studies in genetic mouse models of ASD suggest abnormalities in specific brain regions, as well as in certain cell types. Two studies analyzed the coexpression patterns of a number of ASD genes in the human brain (65, 66). One study found enrichment in mid-gestation layer 5/6 cortical projection neurons, and the other found enrichment in superficial cortical layers and glutamatergic projection neurons. Although the exact layers of the cortex involved were different in the two analyses, the fact that cortical projection neurons were indicated in both studies is potentially important.

In addition to cortical projection neurons, there is increasing evidence for the role of other neuronal subtypes in the pathogenesis of ASD. Multiple mouse models of ASD display reduction in parvalbumin (PV)-cell density in the neocortex (67). PV knockout mice display behavioral phenotypes with relevance to the core symptoms present in human ASD patients (68). In contrast, other groups have reported a selective increase in PV-immunopositive interneurons in the CA1 and CA3 subfields and calretinin-immunopositive neurons in CA1 in patients with ASD (69). Loss of PTEN in mice results in a preferential loss of a different subtype of GABAergic ( $\gamma$ -aminobutyric acid-releasing) neurons, somatostatin (SST) interneurons (70). Interneuron-specific deletions of ASD-related genes result in neurodevelopmental deficits in

### Fig. 2. Hypothetical matrix of genetic mutations and brain regions mapping onto behavioral profiles.

The approach to focus on mechanistic descriptions of symptom clusters rather than symptom inventories requires an understanding of the neural circuit(s) underlying these

behavioral symptoms. One way to examine the neural circuits in animal models is to probe the relationship between a gene's function in a certain brain region and the behavioral deficits in the animal. Use of conditional knockout mice has started to provide such information in certain genetic diseases such as TSC and RTT (93, 118). This matrix represents a hypothetical framework, which needs to be populated by future experimentation. One concrete example of this approach is currently in effect in epilepsy. Absence seizures are thought to arise from voltage-gated calcium channel dysfunction in the thalamus and respond best to ethosuximide treatment. In contrast, complex partial seizures occur as a result of increased excitation or decreased inhibition and thus respond to glutamate antagonists or GABAergic agonists. Such delineation of genetic, cellular, and circuit defects also may prove helpful in treating behavioral deficits associated with ASD with better precision.



mice. For example, loss of MeCP2 from GABAergic interneurons leads to autistic-like repetitive movements, seizures, and deficits in auditory event-related potentials (71, 72). Deficits in inhibitory neurotransmission, along with altered balance of excitation and inhibition (73), have been consistently observed in cortical and hippocampal neurons and circuits in diverse mouse models (74–77). In addition, the reversal potential of GABA may not mature fully when specific ASD genes are mutated, causing GABA to be depolarizing rather than hyperpolarizing (78). Consistent with these findings, a propensity for seizures is a major phenotype of ASDs. Taken together, these findings make a compelling case for dysregulation of inhibition as having a major role in neurodevelopmental disorders. More generally, cell type-specific and brain region-specific deletion of ASD genes is crucial for dissecting the circuit pathophysiology of ASD and in tying it to distinct symptom domains.

Connections between basal ganglia and cortex may underlie certain aspects of ASD. *Neurologin1* knockout mice exhibit ASD-like repetitive behaviors and abnormal corticostriatal synapses (79). *Neurologin3* mutants have similar abnormalities, but the defect appears to be due to a selective synaptic impairment in the nucleus accumbens/ventral striatum (80). SHANK3 is expressed in the basal ganglia, and *Shank3* knockout mice exhibit repetitive grooming behavior, abnormal social interactions, and changes at corticostriatal synapses (81).

The cerebellum is implicated in the pathogenesis of ASD via histopathology, imaging, and

epidemiological studies of injury. First, neuropathological studies demonstrate loss of cerebellar Purkinje cells in individuals diagnosed with ASD versus typically developing controls (82–85). Second, imaging studies of patients diagnosed with ASD indicate gray and white matter abnormalities in the cerebellum, dating to early childhood (86–90). Premature infants with isolated cerebellar hemorrhage have a higher incidence of ASDs, suggesting that cerebellar dysfunction early in life contributes to the pathogenesis of autism (15, 16). The developmental vulnerability of this circuit is further illustrated by study of genetic syndromes associated with ASD. Positron emission tomography studies in pediatric TSC patients with ASD demonstrate hypermetabolism in the cerebellar nuclei—the output of the cerebellar cortex—in TSC patients with ASD but not in TSC patients without ASD (91). The selective loss of *Tsc1* or *Tsc2* genes in the output cells of the cerebellum, the Purkinje neurons, appears to be sufficient to lead to an autistic-like phenotype in the two mouse models of TSC (92, 93). These findings suggest that abnormal cerebellar function contributes to ASD.

Non-neuronal cells in the brain such as astrocytes and microglia have also been implicated in the pathogenesis of neurodevelopmental disorders (94). Astrocyte processes extend into excitatory synapses, and they influence synaptic development (95) and synaptic transmission via uptake of glutamate (96), as well as by calcium-mediated alterations in synaptic function and plasticity (97, 98). ASD genes such as *Fmr1* and *Mecp2* are now known to influence astrocyte function (99).

Astrocytes express mGluRs, providing a pathway for mGluR signaling to influence fragile X pathophysiology (100). Astrocyte-specific restoration of MeCP2 in *Mecp2* mutant mice restores function (101). Microglia also shape neuronal development and plasticity, and modulate synaptic transmission in the adult brain, via cytokine and chemokine release, as well as phagocytosis (102, 103). Transplantation of wild-type microglia has been reported as reversing symptoms in a mouse model of Rett syndrome, though the interpretation of these findings remains controversial (104, 105).

## Treatments

Despite the many discoveries in basic neuroscience and human genetics, FDA-approved drugs for ASD patients are limited to risperidone (a dopamine antagonist) and aripiprazole (a dopamine agonist), which are both aimed at treating irritability and not the core features of ASD. Given the large number of genes that potentially confer ASD risk, the genetic heterogeneity of ASD presents a substantial obstacle to development of one-size-fits-all therapies. One can imagine several scenarios. It would be ideal to have one treatment for all causes of ASD. This seems rather unlikely; ASD is not one disease, and some genetic causes of ASD appear to have diametrically opposite manifestations at the synaptic level (106). It is also equally unlikely that different interventions can be developed for every genetic cause of ASD. So, the most realistic (and hopeful) scenario is that there will be a convergence upon a few molecular and circuit pathways that can be targeted by a limited number of interventions. The current

**Table 1. Clinical trials for genetically defined neurodevelopmental disorders.** The table lists the trials listed for genetic syndromes highly associated with ASD according to a search on the clinicaltrials.gov website on 30 July 2015. Only actively recruiting or recently completed trials are included. The final column lists primary or secondary end points with relevance to neurodevelopment.

Disorder	Study drug	ASD/neurocognitive outcomes
Tuberous sclerosis	Everolimus (RAD001)	ASD, memory, language skills, cognition, general executive function outcomes, behavioral changes, frequency or reduction of epileptiform events, reduced mTOR signaling
	rhIGF-1	Behavior, cognition, cortical function, motor function
Rett syndrome	Fingolimod (FTY720)	Slow regression of motor or language skills
	Dextromethorphan	Seizures, behavioral problems, cognition
	Glatiramer acetate	Epileptic activity, general behavior
	NNZ-2566	EEG, behavior, autonomic function
Fragile X syndrome	Acamprosate	Inattention or hyperactivity, social impairment, behavior, cognition
	NNZ-2566	Behavior, global and functional outcome measures
	Ganaxolone	Behavior, anxiety, attention, cognition
	Metadoxine (MG01CI)	Attention-deficit hyperactivity disorder
	Epigallocatechin-3-gallate (EGCG)	Improve intellectual disability, learning, memory, language
Angelman syndrome	Minocycline	Motor development, behavior, cognition, language
	RG1662	Cognition, behavior
Down syndrome	Low-dose nicotine	Cognitive improvement
	Donepezil (E2020)	Activities of daily living
	Thyroid hormone and folic acid	Psychomotor development
Phelan-McDermid syndrome	rhIGF-1	Behavior, language, motor skills

focus is on the genetic syndromes with high penetrance of ASD symptoms, often caused by single-gene mutations (Table 1). That mouse models of many of the syndromes associated with ASD respond positively to treatment, even in adulthood (107–109), has further bolstered optimism about the utility of pharmacological treatments in these disorders.

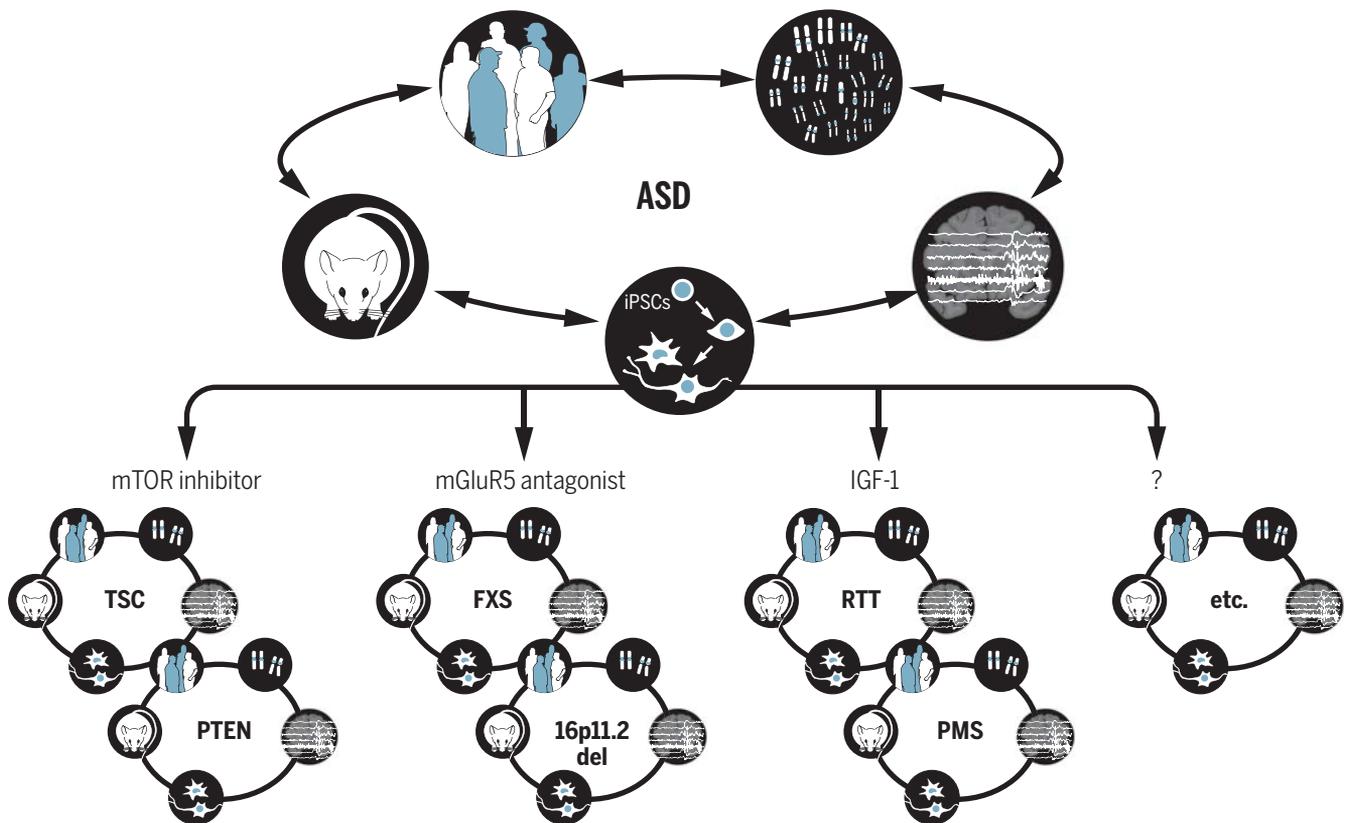
One of the first attempts at testing mechanism-based therapies in ASD was performed in FXS. The mGluR theory of fragile X predicted that many symptoms of FXS are due to exaggerated responses to activation of mGluRs. This was demonstrated to be true in many animal models of FXS (110). Nonetheless, two mGluR antagonists (one made by Roche and another by Novartis) failed to show efficacy in phase 2 trials (111). These negative results highlight the difficulties associated with clinical trials in neurodevelopmental disorders: Did the drugs engage their targets in the central nervous system? Were the end points chosen dynamic within the duration of the trial? Was the placebo effect too large? Was the right group of patients (e.g., patients at an appropriate stage of symptoms, or a subset with a particular genotype) chosen for enrollment?

Another important issue raised by these studies is how to best utilize animal models for developing therapies. Physiological and behavioral analyses in mice have been crucial for advancing our understanding of circuitry underlying social interactions and repetitive behaviors. However, a “good” mouse model needs to have both construct and face validity (112). More important, the circuit that is being analyzed needs to have some direct relevance to outcome measures in humans. Only then can the pharmacological interventions that modulate that circuit be translated effectively from mice to humans.

It is surprising that relatively few pharmacokinetic and pharmacodynamic (PK/PD) relationships are tested in preclinical studies in mouse models of neurodevelopmental disorders. Although the pharmacokinetics will not be the same for a compound in mice and humans, understanding how much of the target needs to be engaged and over what period of time it must be engaged to achieve efficacy is crucial to interpret the preclinical data correctly and translate the findings to clinical studies. To identify the correct target, more detailed preclinical studies will be necessary going forward. In terms of early-stage clinical

trials, many interventions look promising in open-label studies but fail to show efficacy when compared to placebo. Thus, more placebo-controlled phase 2 trials will be needed.

Biomarkers can be crucial for predicting subjects most likely to respond to a test drug, confirming target engagement, and detecting early signals of efficacy. Finding biomarkers that will segregate similarly diagnosed ASD patients into subsets of biologically more homogeneous populations is a critical feature of good clinical trial design. A “stratification biomarker” can be a biochemical measure from patient samples, a structural feature of a human imaging study, or a functional feature of an imaging or electrophysiological study. Aside from stratification, biomarkers can also be helpful in early diagnosis and assessing phenotype and severity, as well as measuring PK/PD in drug studies. Given that ASD represents circuit dysfunction, biomarkers that allow us to examine ASD-related circuits are likely to be most relevant. Especially, translatable biomarkers that can be used in both mouse models and human subjects can be particularly powerful [e.g., EEG, MRI, visual or auditory evoked potentials, or eye-blink conditioning (64, 113–115)]. Similarly,



**Fig. 3. Translational research and clinical trials in ASD.** Translational studies in ASD have gained momentum from genetically defined causes such as FXS, TSC, and RTT. The patients with these disorders are phenotyped in detail by means of advanced imaging and electrophysiology studies, with the aim of identifying potential biomarkers. There are cell-based models (both rodent and human) as well as mouse models of these syndromes, enabling preclinical trials. Together, these efforts have led to clinical trials in some of these disorders. Based on the preclinical trials, the hypothesis is that different etiologies

of ASD will respond to different therapies, such as mTOR inhibitors (TSC and PTEN), mGluR5 antagonists (FXS and 16p11.2 deletion), and IGF-1 (RTT and PMS). Subsets of nonsyndromic ASD patients may also benefit from one of these therapies, but further studies will be required to provide the tools and methods to stratify the individuals with nonsyndromic ASD into treatment groups. It is important to remember that the discovery cycle will likely take more than one round to achieve safe and effective therapies for these disorders.

outcome measures that are circuit-based may be more fruitful in detecting efficacy rather than measuring global functioning. A panel of relevant biomarkers, which together provide a unique profile of a patient, may be a crucial component of precision trial design in the future.

One of the most important questions about treatments is when mechanism-based treatments need to be initiated. Since the behavioral manifestations of ASD occur quite early in development, one may have to intervene before symptoms arise. Animal models of syndromic ASD indicate that restoring function well into adulthood can rescue some of the symptoms of the disease (107–109). It is not yet clear if the same is true in humans and what exactly the critical windows for treatment are. However, regardless of age at treatment onset, relevant biomarkers and efficacy measures would be important for establishing the effectiveness of treatment.

One potential new tool to identify subjects who are likely to respond to a test drug is iPSC-derived neurons. This technology allows the possibility of testing the effects of a compound on a patient's neurons first before giving it to the patient. Modeling the effects of mutations in iPSC-derived neurons can be informative about the molecular and cellular defects but is unlikely with the current technology to provide insights on the emergent dysfunctions at the level of neuronal circuits. Nonetheless, a preliminary testing of efficacy in a patient's iPSC-derived neurons should be a vital component of trial design for precision medicine.

Since many of the diseases being targeted in these initial trials are genetic, they may be amenable to gene therapy in theory. Gene therapy using viral vectors is undergoing a renaissance and may be particularly applicable to diseases that arise from loss of function of a particular gene such as *MECP2* or *CDKL5* (*cyclin-dependent kinase-like 5*). Aside from the delivery issues, one must pay close attention to the dosage effects, because many of the genes that result in an ASD-related phenotype also have deleterious effects when they are expressed at high dosage. Thus, expression of the exogenous genes may have to be regulated tightly in spatial and temporal terms, as well as levels of expression.

If we reach success in single genetically defined syndromes, there will be two new roadblocks in generalizing these findings to the larger ASD population. First, such an extension will require a comparative analysis of the different genetically defined causes of ASD to determine whether effective treatments in one may also be effective in another (Fig. 3). Such comparative understanding of the genetic etiologies underlying ASD is in its infancy (106). A second and more difficult hurdle will be applying these findings to the "idiopathic" genetically unknown or undefined ASD population. Currently, we do not have an analytical tool to determine if an individual with ASD would benefit from a treatment that is effective, for example, in TSC or in FXS. A marker to classify patients according to genetic, biochemical, or circuit abnormalities does not

yet exist. However, even single-gene conditions involve multiple potential targets, and combination therapies are likely to be more effective than single drugs for single targets (47). The success of targeted pharmacological interventions would require integration of multiple kinds of data: knowledge of the genetic mutation and its signaling pathways and synaptic molecules, effectiveness of the therapy on neuronal and synaptic phenotypes in patient-derived neurons and non-neuronal cells in culture, and even analysis of transplanted human neurons in mice.

Although pharmacological treatments may normalize neuronal and synaptic abnormalities, cognitive function is still dependent on complex circuits and interaction of the individual with his or her environment. Thus, pharmacological treatments alone may not be sufficient to reach the optimal outcome without behavioral treatments. Behavioral interventions appear promising in mouse models (116, 117) and could be combined with pharmacological interventions in future clinical trials. While simply correcting the synaptic abnormality with a pharmacological agent may not be sufficient to affect behavioral changes, it could accelerate the rate of learning and sociability in the setting of behavioral interventions. Although combining treatments adds complexity to the trial design, a few such trials are in the planning stages. Trials based on a mechanistic understanding of the disease, performed on a well-defined group of subjects, with evidence of target engagement and supportive biomarkers, are the most likely to succeed. Once such trials prove effective in the highly penetrant genetic syndromes, the next challenge will be to identify patients with idiopathic autism who may benefit from the same treatment. Such an approach will finally realize the notion of precision medicine for autism and related neurodevelopmental disorders.

#### REFERENCES AND NOTES

- L. Kanner, Autistic disturbances of affective contact. *Nerv. Child* **2**, 217–250 (1943).
- C. Lord, in *Understanding Autism: From Basic Neuroscience to Treatment*, S. O. Moldin, J. L. R. Rubenstein, Ed. (Taylor & Francis, Boca Raton, FL, 2006), pp. 1–23.
- M. Ganz, in *Understanding Autism: From Basic Neuroscience to Treatment*, S. O. Moldin, J. L. R. Rubenstein, Ed. (Taylor & Francis, Boca Raton, FL, 2006), pp. 475–502.
- G. Vivanti, J. Barbaro, K. Hudry, C. Dissanayake, M. Prior, Intellectual development in autism spectrum disorders: New insights from longitudinal studies. *Front. Hum. Neurosci.* **7**, 354 (2013). doi: [10.3389/fnhum.2013.00354](https://doi.org/10.3389/fnhum.2013.00354); pmid: [23847518](https://pubmed.ncbi.nlm.nih.gov/23847518/)
- C. Gillberg, E. Fernell, Autism plus versus autism pure. *J. Autism Dev. Disord.* **44**, 3274–3276 (2014). doi: [10.1007/s10803-014-2163-1](https://doi.org/10.1007/s10803-014-2163-1); pmid: [24958434](https://pubmed.ncbi.nlm.nih.gov/24958434/)
- D. Ebrahimi-Fakhari, M. Sahin, Autism and the synapse: Emerging mechanisms and mechanism-based therapies. *Curr. Opin. Neurol.* **28**, 91–102 (2015). doi: [10.1097/WCO.000000000000186](https://doi.org/10.1097/WCO.000000000000186); pmid: [25695134](https://pubmed.ncbi.nlm.nih.gov/25695134/)
- B. J. Casey, M. E. Oliveri, T. Insel, A neurodevelopmental perspective on the research domain criteria (RDoC) framework. *Biol. Psychiatry* **76**, 350–353 (2014). doi: [10.1016/j.biopsych.2014.01.006](https://doi.org/10.1016/j.biopsych.2014.01.006); pmid: [25103538](https://pubmed.ncbi.nlm.nih.gov/25103538/)
- T. Insel et al., Research domain criteria (RDoC): Toward a new classification framework for research on mental disorders. *Am. J. Psychiatry* **167**, 748–751 (2010). doi: [10.1176/appi.ajp.2010.09091379](https://doi.org/10.1176/appi.ajp.2010.09091379); pmid: [20595427](https://pubmed.ncbi.nlm.nih.gov/20595427/)
- D. H. Geschwind, M. W. State, Gene hunting in autism spectrum disorder: On the path to precision medicine. *Lancet Neurol.* (2015). doi: [10.1016/S1474-4422\(15\)00044-7](https://doi.org/10.1016/S1474-4422(15)00044-7); pmid: [25891009](https://pubmed.ncbi.nlm.nih.gov/25891009/)

- S. Ripke et al., Biological insights from 108 schizophrenia-associated genetic loci. *Nature* **511**, 421–427 (2014). doi: [10.1038/nature13595](https://doi.org/10.1038/nature13595); pmid: [25056061](https://pubmed.ncbi.nlm.nih.gov/25056061/)
- A. Poduri, G. D. Evrony, X. Cai, C. A. Walsh, Somatic mutation, genomic variation, and neurological disease. *Science* **341**, 1237758 (2013). doi: [10.1126/science.1237758](https://doi.org/10.1126/science.1237758); pmid: [23828942](https://pubmed.ncbi.nlm.nih.gov/23828942/)
- T. W. Yu et al., Using whole-exome sequencing to identify inherited causes of autism. *Neuron* **77**, 259–273 (2013). pmid: [23352163](https://pubmed.ncbi.nlm.nih.gov/23352163/)
- E. M. Morrow et al., Identifying autism loci and genes by tracing recent shared ancestry. *Science* **321**, 218–223 (2008). doi: [10.1126/science.1157657](https://doi.org/10.1126/science.1157657); pmid: [18621663](https://pubmed.ncbi.nlm.nih.gov/18621663/)
- T. N. Turner et al., Loss of  $\delta$ -catenin function in severe autism. *Nature* **520**, 51–56 (2015). doi: [10.1038/nature14186](https://doi.org/10.1038/nature14186); pmid: [25807484](https://pubmed.ncbi.nlm.nih.gov/25807484/)
- C. Limperopoulos et al., Does cerebellar injury in premature infants contribute to the high prevalence of long-term cognitive, learning, and behavioral disability in survivors? *Pediatrics* **120**, 584–593 (2007). doi: [10.1542/peds.2007-1041](https://doi.org/10.1542/peds.2007-1041); pmid: [17766532](https://pubmed.ncbi.nlm.nih.gov/17766532/)
- C. Limperopoulos, G. Chilingaryan, N. Guizard, R. L. Robertson, A. J. Du Plessis, Cerebellar injury in the premature infant is associated with impaired growth of specific cerebral regions. *Pediatr. Res.* **68**, 145–150 (2010). doi: [10.1203/PDR.0b013e3181e1d032](https://doi.org/10.1203/PDR.0b013e3181e1d032); pmid: [20389260](https://pubmed.ncbi.nlm.nih.gov/20389260/)
- L. Shi, S. H. Fatemi, R. W. Sidwell, P. H. Patterson, Maternal influenza infection causes marked behavioral and pharmacological changes in the offspring. *J. Neurosci.* **23**, 297–302 (2003). pmid: [12514227](https://pubmed.ncbi.nlm.nih.gov/12514227/)
- P. H. Patterson, Immune involvement in schizophrenia and autism: Etiology, pathology and animal models. *Behav. Brain Res.* **204**, 313–321 (2009). doi: [10.1016/j.bbr.2008.12.016](https://doi.org/10.1016/j.bbr.2008.12.016); pmid: [19136031](https://pubmed.ncbi.nlm.nih.gov/19136031/)
- P. A. Garay, A. K. McAllister, Novel roles for immune molecules in neural development: Implications for neurodevelopmental disorders. *Front. Synaptic Neurosci.* **2**, 136 (2010). doi: [10.3389/fnsyn.2010.00136](https://doi.org/10.3389/fnsyn.2010.00136); pmid: [21423522](https://pubmed.ncbi.nlm.nih.gov/21423522/)
- K. Garbett et al., Immune transcriptome alterations in the frontal cortex of subjects with autism. *Neurobiol. Dis.* **30**, 303–311 (2008). doi: [10.1016/j.nbd.2008.01.012](https://doi.org/10.1016/j.nbd.2008.01.012); pmid: [18378158](https://pubmed.ncbi.nlm.nih.gov/18378158/)
- D. L. Vargas, C. Nascimbene, C. Krishnan, A. W. Zimmerman, C. A. Pardo, Neuroglial activation and neuroinflammation in the brain of patients with autism. *Ann. Neurol.* **57**, 67–81 (2005). doi: [10.1002/ana.20315](https://doi.org/10.1002/ana.20315); pmid: [15546155](https://pubmed.ncbi.nlm.nih.gov/15546155/)
- P. A. Garay, E. Y. Hsiao, P. H. Patterson, A. K. McAllister, Maternal immune activation causes age- and region-specific changes in brain cytokines in offspring throughout development. *Brain Behav. Immun.* **31**, 54–68 (2013). doi: [10.1016/j.bbi.2012.07.008](https://doi.org/10.1016/j.bbi.2012.07.008); pmid: [22841693](https://pubmed.ncbi.nlm.nih.gov/22841693/)
- R. K. Weir et al., Preliminary evidence of neuropathology in nonhuman primates prenatally exposed to maternal immune activation. *Brain Behav. Immun.* **48**, 139–146 (2015). doi: [10.1016/j.bbi.2015.03.009](https://doi.org/10.1016/j.bbi.2015.03.009); pmid: [25816799](https://pubmed.ncbi.nlm.nih.gov/25816799/)
- D. Ehninger et al., Gestational immune activation and Tsc2 haploinsufficiency cooperate to disrupt fetal survival and may perturb social behavior in adult mice. *Mol. Psychiatry* **17**, 62–70 (2012). doi: [10.1038/mp.2010.115](https://doi.org/10.1038/mp.2010.115); pmid: [21079609](https://pubmed.ncbi.nlm.nih.gov/21079609/)
- J. G. Mulle, W. G. Sharp, J. F. Cubells, The gut microbiome: A new frontier in autism research. *Curr. Psychiatry Rep.* **15**, 337 (2013). doi: [10.1007/s11920-012-0337-0](https://doi.org/10.1007/s11920-012-0337-0); pmid: [23307560](https://pubmed.ncbi.nlm.nih.gov/23307560/)
- D. W. Kang et al., Reduced incidence of Prevotella and other fermenters in intestinal microflora of autistic children. *PLoS ONE* **8**, e68322 (2013). pmid: [23844187](https://pubmed.ncbi.nlm.nih.gov/23844187/)
- E. Y. Hsiao et al., Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell* **155**, 1451–1463 (2013). doi: [10.1016/j.cell.2013.11.024](https://doi.org/10.1016/j.cell.2013.11.024); pmid: [24315484](https://pubmed.ncbi.nlm.nih.gov/24315484/)
- R. J. Kelleher 3rd, M. F. Bear, The autistic neuron: Troubled translation? *Cell* **135**, 401–406 (2008). doi: [10.1016/j.cell.2008.10.017](https://doi.org/10.1016/j.cell.2008.10.017); pmid: [18984149](https://pubmed.ncbi.nlm.nih.gov/18984149/)
- H. Y. Zoghbi, M. F. Bear, Synaptic dysfunction in neurodevelopmental disorders associated with autism and intellectual disabilities. *Cold Spring Harb. Perspect. Biol.* **4**, a009886 (2012). doi: [10.1101/cshperspect.a009886](https://doi.org/10.1101/cshperspect.a009886); pmid: [22258914](https://pubmed.ncbi.nlm.nih.gov/22258914/)
- T. C. Südhof, Neurotrogins and neuroligins link synaptic function to cognitive disease. *Nature* **455**, 903–911 (2008). doi: [10.1038/nature07456](https://doi.org/10.1038/nature07456); pmid: [18923512](https://pubmed.ncbi.nlm.nih.gov/18923512/)
- I. lossifov et al., De novo gene disruptions in children on the autistic spectrum. *Neuron* **74**, 285–299 (2012). doi: [10.1016/j.neuron.2012.04.009](https://doi.org/10.1016/j.neuron.2012.04.009); pmid: [22542183](https://pubmed.ncbi.nlm.nih.gov/22542183/)

32. R. Bernier *et al.*, Disruptive CHD8 mutations define a subtype of autism early in development. *Cell* **158**, 263–276 (2014). doi: [10.1016/j.cell.2014.06.017](https://doi.org/10.1016/j.cell.2014.06.017); pmid: 24998929
33. J. Castro, N. Mellios, M. Sur, Mechanisms and therapeutic challenges in autism spectrum disorders: Insights from Rett syndrome. *Curr. Opin. Neurol.* **26**, 154–159 (2013). doi: [10.1097/WCO.0b013e32835f19a7](https://doi.org/10.1097/WCO.0b013e32835f19a7); pmid: 23449173
34. X. Nan, F. J. Campoy, A. Bird, MeCP2 is a transcriptional repressor with abundant binding sites in genomic chromatin. *Cell* **88**, 471–481 (1997). doi: [10.1016/S0092-8674\(00\)81887-5](https://doi.org/10.1016/S0092-8674(00)81887-5); pmid: 9038338
35. R. R. Meehan, J. D. Lewis, A. P. Bird, Characterization of MeCP2, a vertebrate DNA binding protein with affinity for methylated DNA. *Nucleic Acids Res.* **20**, 5085–5092 (1992). doi: [10.1093/nar/20.19.5085](https://doi.org/10.1093/nar/20.19.5085); pmid: 1408825
36. P. L. Jones *et al.*, Methylated DNA and MeCP2 recruit histone deacetylase to repress transcription. *Nat. Genet.* **19**, 187–191 (1998). doi: [10.1038/561](https://doi.org/10.1038/561); pmid: 9620779
37. M. Chahrouh *et al.*, MeCP2, a key contributor to neurological disease, activates and represses transcription. *Science* **320**, 1224–1229 (2008). doi: [10.1126/science.1153252](https://doi.org/10.1126/science.1153252); pmid: 18511691
38. S. Ben-Shachar, M. Chahrouh, C. Thaller, C. A. Shaw, H. Y. Zoghbi, Mouse models of MeCP2 disorders share gene expression changes in the cerebellum and hypothalamus. *Hum. Mol. Genet.* **18**, 2431–2442 (2009). doi: [10.1093/hmg/ddp181](https://doi.org/10.1093/hmg/ddp181); pmid: 19369296
39. R. G. Urdinguio *et al.*, Disrupted microRNA expression caused by MeCP2 loss in a mouse model of Rett syndrome. *Epigenetics* **5**, 656–663 (2010). doi: [10.4161/epi.5.7.13055](https://doi.org/10.4161/epi.5.7.13055); pmid: 20716963
40. H. Wu *et al.*, Genome-wide analysis reveals methyl-CpG-binding protein 2-dependent regulation of microRNAs in a mouse model of Rett syndrome. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 18161–18166 (2010). doi: [10.1073/pnas.1005595107](https://doi.org/10.1073/pnas.1005595107); pmid: 20921386
41. P. Jin *et al.*, Biochemical and genetic interaction between the fragile X mental retardation protein and the microRNA pathway. *Nat. Neurosci.* **7**, 113–117 (2004). doi: [10.1038/nrn1174](https://doi.org/10.1038/nrn1174); pmid: 14703574
42. J. O. Lipton, M. Sahin, The neurology of mTOR. *Neuron* **84**, 275–291 (2014). doi: [10.1016/j.neuron.2014.09.034](https://doi.org/10.1016/j.neuron.2014.09.034); pmid: 25374355
43. M. Qin *et al.*, Altered cerebral protein synthesis in fragile X syndrome: Studies in human subjects and knockout mice. *J. Cereb. Blood Flow Metab.* **33**, 499–507 (2013). doi: [10.1038/jcbfm.2012.205](https://doi.org/10.1038/jcbfm.2012.205); pmid: 23299245
44. J. C. Darnell *et al.*, FMRP stalls ribosomal translocation on mRNAs linked to synaptic function and autism. *Cell* **146**, 247–261 (2011). doi: [10.1016/j.cell.2011.06.013](https://doi.org/10.1016/j.cell.2011.06.013); pmid: 21784246
45. Q. Chang, G. Khare, V. Dani, S. Nelson, R. Jaenisch, The disease progression of MeCP2 mutant mice is affected by the level of BDNF expression. *Neuron* **49**, 341–348 (2006). doi: [10.1016/j.neuron.2005.12.027](https://doi.org/10.1016/j.neuron.2005.12.027); pmid: 16446138
46. D. Tropea *et al.*, Partial reversal of Rett Syndrome-like symptoms in MeCP2 mutant mice. *Proc. Natl. Acad. Sci. U.S.A.* **106**, 2029–2034 (2009). doi: [10.1073/pnas.0812394106](https://doi.org/10.1073/pnas.0812394106); pmid: 19208815
47. N. Mellios *et al.*,  $\beta_2$ -Adrenergic receptor agonist ameliorates phenotypes and corrects microRNA-mediated IGF1 deficits in a mouse model of Rett syndrome. *Proc. Natl. Acad. Sci. U.S.A.* **111**, 9947–9952 (2014). doi: [10.1073/pnas.1309426111](https://doi.org/10.1073/pnas.1309426111); pmid: 24958851
48. J. Castro *et al.*, Functional recovery with recombinant human IGF1 treatment in a mouse model of Rett Syndrome. *Proc. Natl. Acad. Sci. U.S.A.* **111**, 9941–9946 (2014). doi: [10.1073/pnas.1311685111](https://doi.org/10.1073/pnas.1311685111); pmid: 24958891
49. S. Ricciardi *et al.*, Reduced AKT/mTOR signaling and protein synthesis dysregulation in a Rett syndrome animal model. *Hum. Mol. Genet.* **20**, 1182–1196 (2011). doi: [10.1093/hmg/ddq563](https://doi.org/10.1093/hmg/ddq563); pmid: 21212100
50. M. C. Marchetto *et al.*, A model for neural development and treatment of Rett syndrome using human induced pluripotent stem cells. *Cell* **143**, 527–539 (2010). doi: [10.1016/j.cell.2010.10.016](https://doi.org/10.1016/j.cell.2010.10.016); pmid: 21074045
51. Y. Li *et al.*, Global transcriptional and translational repression in human-embryonic-stem-cell-derived Rett syndrome neurons. *Cell Stem Cell* **13**, 446–458 (2013). doi: [10.1016/j.stem.2013.09.001](https://doi.org/10.1016/j.stem.2013.09.001); pmid: 24094325
52. O. S. Khwaja *et al.*, Safety, pharmacokinetics, and preliminary assessment of efficacy of mecamsermin (recombinant human IGF-1) for the treatment of Rett syndrome. *Proc. Natl. Acad. Sci. U.S.A.* **111**, 4596–4601 (2014). doi: [10.1073/pnas.1311411111](https://doi.org/10.1073/pnas.1311411111); pmid: 24623853
53. A. Shcheglovitov *et al.*, SHANK3 and IGF1 restore synaptic deficits in neurons from 22q13 deletion syndrome patients. *Nature* **503**, 267–271 (2013). pmid: 24132240
54. O. Bozdagi, T. Tavassoli, J. D. Buxbaum, Insulin-like growth factor-1 rescues synaptic and motor deficits in a mouse model of autism and developmental delay. *Mol. Autism* **4**, 9 (2013). doi: [10.1186/2040-2392-4-9](https://doi.org/10.1186/2040-2392-4-9); pmid: 23621888
55. A. Bhattacharya *et al.*, Genetic removal of p70 S6 kinase 1 corrects molecular, synaptic, and behavioral phenotypes in fragile X syndrome mice. *Neuron* **76**, 325–337 (2012). doi: [10.1016/j.neuron.2012.07.022](https://doi.org/10.1016/j.neuron.2012.07.022); pmid: 23083736
56. T. Udagawa *et al.*, Genetic and acute CPEB1 depletion ameliorate fragile X pathophysiology. *Nat. Med.* **19**, 1473–1477 (2013). doi: [10.1038/nm.3353](https://doi.org/10.1038/nm.3353); pmid: 2441422
57. C. Gross *et al.*, Selective role of the catalytic PI3K subunit p110 $\beta$  in impaired higher order cognition in fragile X syndrome. *Cell Rep.* **11**, 681–688 (2015). doi: [10.1016/j.celrep.2015.03.065](https://doi.org/10.1016/j.celrep.2015.03.065); pmid: 25921527
58. C. Gross *et al.*, Increased expression of the PI3K enhancer PIKE mediates deficits in synaptic plasticity and behavior in fragile X syndrome. *Cell Rep.* **11**, 727–736 (2015). doi: [10.1016/j.celrep.2015.03.060](https://doi.org/10.1016/j.celrep.2015.03.060); pmid: 25921541
59. K. M. Huber, M. S. Kayser, M. F. Bear, Role for rapid dendritic protein synthesis in hippocampal mGluR-dependent long-term depression. *Science* **288**, 1254–1256 (2000). doi: [10.1126/science.288.5469.1254](https://doi.org/10.1126/science.288.5469.1254); pmid: 10818003
60. K. M. Huber, S. M. Gallagher, S. T. Warren, M. F. Bear, Altered synaptic plasticity in a mouse model of fragile X mental retardation. *Proc. Natl. Acad. Sci. U.S.A.* **99**, 7746–7750 (2002). doi: [10.1073/pnas.122205699](https://doi.org/10.1073/pnas.122205699); pmid: 12032354
61. D. Tian *et al.*, Contribution of mGluR5 to pathophysiology in a mouse model of human chromosome 16p11.2 microdeletion. *Nat. Neurosci.* **18**, 182–184 (2015). doi: [10.1038/nrn.3911](https://doi.org/10.1038/nrn.3911); pmid: 25581360
62. S. J. Baudouin *et al.*, Shared synaptic pathophysiology in syndromic and nonsyndromic rodent models of autism. *Science* **338**, 128–132 (2012). doi: [10.1126/science.1224159](https://doi.org/10.1126/science.1224159); pmid: 22983708
63. A. A. Scott-Van Zeeland *et al.*, Altered functional connectivity in frontal lobe circuits is associated with variation in the autism risk gene CNTNAP2. *Sci. Transl. Med.* **2**, 56ra80 (2010). doi: [10.1126/scitranslmed.3001344](https://doi.org/10.1126/scitranslmed.3001344); pmid: 21048216
64. J. M. Peters *et al.*, Loss of white matter microstructural integrity is associated with adverse neurological outcome in tuberous sclerosis complex. *Acad. Radiol.* **19**, 17–25 (2012). doi: [10.1016/j.acra.2011.08.016](https://doi.org/10.1016/j.acra.2011.08.016); pmid: 22142677
65. A. J. Willsey *et al.*, Coexpression networks implicate human midfetal deep cortical projection neurons in the pathogenesis of autism. *Cell* **155**, 997–1007 (2013). doi: [10.1016/j.cell.2013.10.020](https://doi.org/10.1016/j.cell.2013.10.020); pmid: 24267886
66. N. N. Parikshak *et al.*, Integrative functional genomic analyses implicate specific molecular pathways and circuits in autism. *Cell* **155**, 1008–1021 (2013). doi: [10.1016/j.cell.2013.10.031](https://doi.org/10.1016/j.cell.2013.10.031); pmid: 24267887
67. N. Gogolla *et al.*, Common circuit defect of excitatory-inhibitory balance in mouse models of autism. *J. Neurodev. Disord.* **1**, 172–181 (2009). doi: [10.1007/s11689-009-9023-x](https://doi.org/10.1007/s11689-009-9023-x); pmid: 20664807
68. M. Wöhr *et al.*, Lack of parvalbumin in mice leads to behavioral deficits relevant to all human autism core symptoms and related neural morphofunctional abnormalities. *Transl. Psychiatry* **5**, e525 (2015). doi: [10.1038/tp.2015.19](https://doi.org/10.1038/tp.2015.19); pmid: 25756808
69. Y. A. Lawrence, T. L. Kemper, M. L. Bauman, G. J. Blatt, Parvalbumin-, calbindin-, and calretinin-immunoreactive hippocampal interneuron density in autism. *Acta Neurol. Scand.* **121**, 99–108 (2010). doi: [10.1111/j.1600-0404.2009.01234.x](https://doi.org/10.1111/j.1600-0404.2009.01234.x); pmid: 19719810
70. D. Vogt, K. K. Cho, A. T. Lee, V. S. Sohal, J. L. Rubenstein, The parvalbumin/somatostatin ratio is increased in Pten mutant mice and by human PTEN ASD alleles. *Cell Rep.* **11**, 944–956 (2015). doi: [10.1016/j.celrep.2015.04.019](https://doi.org/10.1016/j.celrep.2015.04.019); pmid: 25937288
71. D. Goffin, E. S. Brodwin, J. A. Blendy, S. J. Siegel, Z. Zhou, Cellular origins of auditory event-related potential deficits in Rett syndrome. *Nat. Neurosci.* **17**, 804–806 (2014). doi: [10.1038/nrn.3710](https://doi.org/10.1038/nrn.3710); pmid: 24777420
72. H. T. Chao *et al.*, Dysfunction in GABA signalling mediates autism-like stereotypies and Rett syndrome phenotypes. *Nature* **468**, 263–269 (2010). doi: [10.1038/nature09582](https://doi.org/10.1038/nature09582); pmid: 21068835
73. J. L. Rubenstein, M. M. Merzenich, Model of autism: Increased ratio of excitation/inhibition in key neural systems. *Genes Brain Behav.* **2**, 255–267 (2003). doi: [10.1034/j.1601-183X.2003.00037.x](https://doi.org/10.1034/j.1601-183X.2003.00037.x); pmid: 14606691
74. G. Calfa, W. Li, J. M. Rutherford, L. Pozzo-Miller, Excitation/inhibition imbalance and impaired synaptic inhibition in hippocampal area CA3 of MeCP2 knockout mice. *Hippocampus* **25**, 159–168 (2015). doi: [10.1002/hipo.22360](https://doi.org/10.1002/hipo.22360); pmid: 25209930
75. V. S. Dani *et al.*, Reduced cortical activity due to a shift in the balance between excitation and inhibition in a mouse model of Rett syndrome. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 12560–12565 (2005). doi: [10.1073/pnas.0506071102](https://doi.org/10.1073/pnas.0506071102); pmid: 16116096
76. H. T. Chao, H. Y. Zoghbi, C. Rosenmund, MeCP2 controls excitatory synaptic strength by regulating glutamatergic synapse number. *Neuron* **56**, 58–65 (2007). doi: [10.1016/j.neuron.2007.08.018](https://doi.org/10.1016/j.neuron.2007.08.018); pmid: 17920015
77. L. Wood, N. W. Gray, Z. Zhou, M. E. Greenberg, G. M. Shepherd, Synaptic circuit abnormalities of motor-frontal layer 2/3 pyramidal neurons in an RNA interference model of methyl-CpG-binding protein 2 deficiency. *J. Neurosci.* **29**, 12440–12448 (2009). doi: [10.1523/JNEUROSCI.3321-09.2009](https://doi.org/10.1523/JNEUROSCI.3321-09.2009); pmid: 19812320
78. Y. Ben-Ari, J. L. Gaiarsa, R. Tyzio, R. Khazipov, GABA: A pioneer transmitter that excites immature neurons and generates primitive oscillations. *Physiol. Rev.* **87**, 1215–1284 (2007). doi: [10.1152/physrev.00017.2006](https://doi.org/10.1152/physrev.00017.2006); pmid: 17928584
79. J. Blundell *et al.*, Neuroigin-1 deletion results in impaired spatial memory and increased repetitive behavior. *J. Neurosci.* **30**, 2115–2129 (2010). doi: [10.1523/JNEUROSCI.4517-09.2010](https://doi.org/10.1523/JNEUROSCI.4517-09.2010); pmid: 20147539
80. P. E. Rothwell *et al.*, Autism-associated neuroigin-3 mutations commonly impair striatal circuits to boost repetitive behaviors. *Cell* **158**, 198–212 (2014). doi: [10.1016/j.cell.2014.04.045](https://doi.org/10.1016/j.cell.2014.04.045); pmid: 24995986
81. J. Peça *et al.*, Shank3 mutant mice display autistic-like behaviours and striatal dysfunction. *Nature* **472**, 437–442 (2011). doi: [10.1038/nature09965](https://doi.org/10.1038/nature09965); pmid: 21423165
82. M. L. Bauman, T. L. Kemper, Neuroanatomic observations of the brain in autism: A review and future directions. *Int. J. Dev. Neurosci.* **23**, 183–187 (2005). doi: [10.1016/j.jdevneu.2004.09.006](https://doi.org/10.1016/j.jdevneu.2004.09.006); pmid: 15749244
83. M. F. Casanova, The neuropathology of autism. *Brain Pathol.* **17**, 422–433 (2007). doi: [10.1111/j.1750-3639.2007.00100.x](https://doi.org/10.1111/j.1750-3639.2007.00100.x); pmid: 17919128
84. S. H. Fatemi *et al.*, Consensus paper: Pathological role of the cerebellum in autism. *Cerebellum* **11**, 777–807 (2012). pmid: 22370873
85. E. R. Whitney, T. L. Kemper, M. L. Bauman, D. L. Rosene, G. J. Blatt, Cerebellar Purkinje cells are reduced in a subpopulation of autistic brains: A stereological experiment using calbindin-D28k. *Cerebellum* **7**, 406–416 (2008). doi: [10.1007/s12311-008-0043-y](https://doi.org/10.1007/s12311-008-0043-y); pmid: 18587625
86. E. Courchesne, R. Yeung-Courchesne, G. A. Press, J. L. Hesselink, T. L. Jernigan, Hypoplasia of cerebellar vermal lobules VI and VII in autism. *N. Engl. J. Med.* **318**, 1349–1354 (1988). doi: [10.1056/NEJM198805263182102](https://doi.org/10.1056/NEJM198805263182102); pmid: 3367935
87. D. G. Amaral, C. M. Schumann, C. W. Nordahl, Neuroanatomy of autism. *Trends Neurosci.* **31**, 137–145 (2008). doi: [10.1016/j.tins.2007.12.005](https://doi.org/10.1016/j.tins.2007.12.005); pmid: 18258309
88. W. R. Kates *et al.*, Neuroanatomic variation in monozygotic twin pairs discordant for the narrow phenotype for autism. *Am. J. Psychiatry* **161**, 539–546 (2004). doi: [10.1176/appi.appi.161.3.539](https://doi.org/10.1176/appi.appi.161.3.539); pmid: 14992981
89. N. Akshoomoff *et al.*, Outcome classification of preschool children with autism spectrum disorders using MRI brain measures. *J. Am. Acad. Child Adolesc. Psychiatry* **43**, 349–357 (2004). doi: [10.1097/00004583-200403000-00018](https://doi.org/10.1097/00004583-200403000-00018); pmid: 15076269
90. G. Allen, E. Courchesne, Differential effects of developmental cerebellar abnormality on cognitive and motor functions in the cerebellum: An fMRI study of autism. *Am. J. Psychiatry* **160**, 262–273 (2003). doi: [10.1176/appi.appi.160.2.262](https://doi.org/10.1176/appi.appi.160.2.262); pmid: 12562572
91. E. Asano *et al.*, Autism in tuberous sclerosis complex is related to both cortical and subcortical dysfunction. *Neurology* **57**, 1269–1277 (2001). doi: [10.1212/WNL.57.7.1269](https://doi.org/10.1212/WNL.57.7.1269); pmid: 11591847
92. R. M. Reith *et al.*, Loss of Tsc2 in Purkinje cells is associated with autistic-like behavior in a mouse model of tuberous sclerosis complex. *Neurobiol. Dis.* **51**, 93–103 (2013). doi: [10.1016/j.nbd.2012.10.014](https://doi.org/10.1016/j.nbd.2012.10.014); pmid: 23123587

93. P. T. Tsai *et al.*, Autistic-like behaviour and cerebellar dysfunction in Purkinje cell Tsc1 mutant mice. *Nature* **488**, 647–651 (2012). doi: [10.1038/nature11310](https://doi.org/10.1038/nature11310); pmid: 22763451
94. A. V. Molofsky *et al.*, Astrocytes and disease: A neurodevelopmental perspective. *Genes Dev.* **26**, 891–907 (2012). doi: [10.1101/gad.188326.112](https://doi.org/10.1101/gad.188326.112); pmid: 22549954
95. C. Eroglu, B. A. Barres, Regulation of synaptic connectivity by glia. *Nature* **468**, 223–231 (2010). doi: [10.1038/nature09612](https://doi.org/10.1038/nature09612); pmid: 21068831
96. J. Schummers, H. Yu, M. Sur, Tuned responses of astrocytes and their influence on hemodynamic signals in the visual cortex. *Science* **320**, 1638–1643 (2008). doi: [10.1126/science.1156120](https://doi.org/10.1126/science.1156120); pmid: 18566287
97. N. Chen *et al.*, Nucleus basalis-enabled stimulus-specific plasticity in the visual cortex is mediated by astrocytes. *Proc. Natl. Acad. Sci. U.S.A.* **109**, E2832–E2841 (2012). doi: [10.1073/pnas.1206557109](https://doi.org/10.1073/pnas.1206557109); pmid: 23012414
98. P. G. Haydon, M. Nedergaard, How do astrocytes participate in neural plasticity? *Cold Spring Harb. Perspect. Biol.* **7**, a020438 (2015). doi: [10.1101/cshperspect.a020438](https://doi.org/10.1101/cshperspect.a020438); pmid: 25502516
99. D. H. Yasui *et al.*, MeCP2 modulates gene expression pathways in astrocytes. *Mol. Autism* **4**, 3 (2013). doi: [10.1186/2040-2392-4-3](https://doi.org/10.1186/2040-2392-4-3); pmid: 23351786
100. H. Higashimori *et al.*, Astroglial FMRP-dependent translational down-regulation of mGluR5 underlies glutamate transporter GLT1 dysregulation in the fragile X mouse. *Hum. Mol. Genet.* **22**, 2041–2054 (2013). doi: [10.1093/hmg/ddt055](https://doi.org/10.1093/hmg/ddt055); pmid: 23396537
101. D. T. Liroy *et al.*, A role for glia in the progression of Rett's syndrome. *Nature* **475**, 497–500 (2011). doi: [10.1038/nature10214](https://doi.org/10.1038/nature10214); pmid: 21716289
102. A. L. Xavier, J. R. Menezes, S. A. Goldman, M. Nedergaard, Fine-tuning the central nervous system: Microglial modelling of cells and synapses. *Philos. Trans. R. Soc. London B Biol. Sci.* **369**, 20130593 (2014). doi: [10.1098/rstb.2013.0593](https://doi.org/10.1098/rstb.2013.0593); pmid: 25225087
103. D. P. Schafer, E. K. Lehrman, B. Stevens, The “quad-partite” synapse: Microglia-synapse interactions in the developing and mature CNS. *Glia* **61**, 24–36 (2013). doi: [10.1002/glia.22389](https://doi.org/10.1002/glia.22389); pmid: 22829357
104. N. C. Derecki *et al.*, Wild-type microglia arrest pathology in a mouse model of Rett syndrome. *Nature* **484**, 105–109 (2012). doi: [10.1038/nature10907](https://doi.org/10.1038/nature10907); pmid: 22425995
105. J. Wang *et al.*, Wild-type microglia do not reverse pathology in mouse models of Rett syndrome. *Nature* **521**, E1–E4 (2015). doi: [10.1038/nature14444](https://doi.org/10.1038/nature14444); pmid: 25993969
106. B. D. Auerbach, E. K. Osterweil, M. F. Bear, Mutations causing syndromic autism define an axis of synaptic pathophysiology. *Nature* **480**, 63–68 (2011). doi: [10.1038/nature10658](https://doi.org/10.1038/nature10658); pmid: 22113615
107. J. Guy, J. Gan, J. Selfridge, S. Cobb, A. Bird, Reversal of neurological defects in a mouse model of Rett syndrome. *Science* **315**, 1143–1147 (2007). doi: [10.1126/science.1138389](https://doi.org/10.1126/science.1138389); pmid: 17289941
108. A. Michalon *et al.*, Chronic pharmacological mGlu5 inhibition corrects fragile X in adult mice. *Neuron* **74**, 49–56 (2012). doi: [10.1016/j.neuron.2012.03.009](https://doi.org/10.1016/j.neuron.2012.03.009); pmid: 22500629
109. D. Ehninger *et al.*, Reversal of learning deficits in a Tsc2+/- mouse model of tuberous sclerosis. *Nat. Med.* **14**, 843–848 (2008). doi: [10.1038/nm1788](https://doi.org/10.1038/nm1788); pmid: 18568033
110. D. D. Krueger, M. F. Bear, Toward fulfilling the promise of molecular medicine in fragile X syndrome. *Annu. Rev. Med.* **62**, 411–429 (2011). doi: [10.1146/annurev-med-061109-134644](https://doi.org/10.1146/annurev-med-061109-134644); pmid: 21090964
111. S. H. Scharf, G. Jaeschke, J. G. Wettstein, L. Lindemann, Metabotropic glutamate receptor 5 as drug target for Fragile X syndrome. *Curr. Opin. Pharmacol.* **20**, 124–134 (2015). doi: [10.1016/j.coph.2014.11.004](https://doi.org/10.1016/j.coph.2014.11.004); pmid: 25488569
112. J. L. Silverman, M. Yang, C. Lord, J. N. Crawley, Behavioural phenotyping assays for mouse models of autism. *Nat. Rev. Neurosci.* **11**, 490–502 (2010). doi: [10.1038/nrn2851](https://doi.org/10.1038/nrn2851); pmid: 20559336
113. B. C. Reeb-Sutherland, N. A. Fox, Eyeblink conditioning: A non-invasive biomarker for neurodevelopmental disorders. *J. Autism Dev. Disord.* **45**, 376–394 (2015). doi: [10.1007/s10803-013-1905-9](https://doi.org/10.1007/s10803-013-1905-9); pmid: 23942847
114. M. J. Gandal *et al.*, Validating  $\gamma$  oscillations and delayed auditory responses as translational biomarkers of autism. *Biol. Psychiatry* **68**, 1100–1106 (2010). doi: [10.1016/j.biopsych.2010.09.031](https://doi.org/10.1016/j.biopsych.2010.09.031); pmid: 21130222
115. S. Durand *et al.*, NMDA receptor regulation prevents regression of visual cortical function in the absence of MeCP2. *Neuron* **76**, 1078–1090 (2012). doi: [10.1016/j.neuron.2012.12.004](https://doi.org/10.1016/j.neuron.2012.12.004); pmid: 23259945
116. M. Kondo *et al.*, Environmental enrichment ameliorates a motor coordination deficit in a mouse model of Rett syndrome—MeCP2 gene dosage effects and BDNF expression. *Eur. J. Neurosci.* **27**, 3342–3350 (2008). doi: [10.1111/j.1460-9568.2008.06305.x](https://doi.org/10.1111/j.1460-9568.2008.06305.x); pmid: 18557922
117. L. Restivo *et al.*, Enriched environment promotes behavioral and morphological recovery in a mouse model for the fragile X syndrome. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 11557–11562 (2005). doi: [10.1073/pnas.0504984102](https://doi.org/10.1073/pnas.0504984102); pmid: 16076950
118. S. L. Fyffe *et al.*, Deletion of MeCP2 in Sim1-expressing neurons reveals a critical role for MeCP2 in feeding behavior, aggression, and the response to stress. *Neuron* **59**, 947–958 (2008). doi: [10.1016/j.neuron.2008.07.030](https://doi.org/10.1016/j.neuron.2008.07.030); pmid: 18817733

#### ACKNOWLEDGMENTS

We thank R. Kleiman, A. Poduri, and K. Dies for critically reviewing the manuscript. Due to limited space, we have not quoted all literature in the field, and we apologize to those whose articles are not referenced. Research in M. Sur's laboratory is supported by NIH grants MH085802, NS090473, and EY007023; NSF grant EF1451125; and the Simons Foundation Autism Research Initiative. Research in M. Sahin's laboratory is supported by the NIH (U01 NS082320, P20 NS080199, and P30 HD018655), Department of Defense (W81XWH-13-1-0040), Tuberous Sclerosis Alliance, Autism Speaks, Nancy Lurie Marks Family Foundation, Simons Foundation, Boston Children's Hospital Translational Research Program, and Novartis and Shire. The Developmental Synaptopathies Consortium (U54NS092090) is a part of the National Center for Advancing Translational Sciences (NCATS) Rare Diseases Clinical Research Network (RDCRN). RDCRN is an initiative of the Office of Rare Diseases Research (ORDR), NCATS, funded through collaboration between NCATS, NIMH, the National Institute of Neurological Disorders and Stroke, and the Eunice Kennedy Shriver National Institute of Child Health and Human Development.

10.1126/science.aab3897



**Genes, circuits, and precision therapies for autism and related neurodevelopmental disorders**  
Mustafa Sahin and Mriganka Sur  
*Science* **350**, (2015);  
DOI: 10.1126/science.aab3897

---

*This copy is for your personal, non-commercial use only.*

---

**If you wish to distribute this article to others**, you can order high-quality copies for your colleagues, clients, or customers by [clicking here](#).

**Permission to republish or repurpose articles or portions of articles** can be obtained by following the guidelines [here](#).

**The following resources related to this article are available online at [www.sciencemag.org](http://www.sciencemag.org) (this information is current as of November 21, 2015 ):**

**Updated information and services**, including high-resolution figures, can be found in the online version of this article at:

<http://www.sciencemag.org/content/350/6263/aab3897.full.html>

This article **cites 116 articles**, 30 of which can be accessed free:

<http://www.sciencemag.org/content/350/6263/aab3897.full.html#ref-list-1>